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(54) Title: MAMMALIAN CHEMOKINES; RECEPTORS; REAGENTS; USES

(57) Abstract

Novel chemokines and 7 transmembrane receptors from mammals, reagents related thereto, including purified proteins, specific antibodies, and nucleic acids encoding said chemokines or receptors. Methods of using said reagents and diagnostic kits are also provided.

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MAMMALIAN CHEMOKINES; RECEPTORS; REAGENTS; USES

The present filing claims priority to U.S. Patent Application No. 60/036,715, filed January 23, 1997, which is incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates to compositions related to proteins which function in controlling physiology, development, and/or differentiation of mammalian cells. In particular, it provides proteins which are implicated in the regulation of physiology, development, differentiation, or function of various cell types, e.g., chemokines, 7 transmembrane receptors, reagents related to each, e.g., antibodies or nucleic acids encoding them, and uses thereof.

BACKGROUND OF THE INVENTION

The circulating component of the mammalian circulatory system comprises various cell types, including red and white blood cells of the erythroid and myeloid cell lineages. See, e.g., Rapaport (1987) Introduction to Hematology (2d ed.) Lippincott, Philadelphia, PA; Jandl (1987) Blood: Textbook of Hematology, Little, Brown and Co., Boston, MA.; and Paul (ed.) (1993) Fundamental Immunology (3d ed.) Raven Press, N.Y.

For some time, it has been known that the mammalian immune response is based on a series of complex cellular interactions, called the "immune network." Recent research has provided new insights into the inner workings of this network. While it remains clear that much of the response does, in fact, revolve around the network-like interactions of lymphocytes, macrophages, granulocytes, and other cells, immunologists now generally hold the opinion that soluble proteins, known as lymphokines, cytokines, or monokines, play a critical role in controlling these cellular interactions. Thus, there is considerable interest in the isolation, characterization, and mechanisms of action

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of cell modulatory factors, an understanding of which should lead to significant advancements in the diagnosis and therapy of numerous medical abnormalities, e.g., immune system and other disorders.

Lymphokines apparently mediate cellular activities in a variety of ways. They have been shown to support the proliferation, growth, and differentiation of the pluripotential hematopoietic stem cells into vast numbers of progenitors comprising diverse cellular lineages making up a complex immune system. These interactions between the cellular components are necessary for a healthy immune response. These different cellular lineages often respond in a different manner when lymphokines are administered in conjunction with other agents.

The chemokines are a large and diverse superfamily of proteins. The superfamily is subdivided into two classical branches, based upon whether the first two cysteines in the chemokine motif are adjacent (termed the "C-C" branch), or spaced by an intervening residue ("C-X-C"). A more recently identified branch of chemokines lacks two cysteines in the corresponding motif, and is represented by the chemokines known as lymphotactins. Another recently identified branch has three intervening residues between the two cysteines, e.g., CX3C chemokines. See, e.g., Schall and Bacon (1994) Current Opinion in Immunology 6:865-873; and Bacon and Schall (1996) Int. Arch. Allergy & Immunol. 109:97-109.

The chemokine receptors are typically members of the superfamily of G-protein coupled (or linked) receptors (GPCR, or GPLR). As a class, these receptors are integral membrane proteins characterized by amino acid sequences which contain seven hydrophobic domains. See, e.g., Ruffolo and Hollinger (eds. 1995) G-Protein Coupled Transmembrane Signaling Mechanisms CRC Press, Boca Raton, FL; Watson and Arkinstall (1994) The G-Protein Linked Receptor FactsBook Academic Press, San Diego, CA; Peroutka (ed. 1994) G Protein-Coupled Receptors CRC Press, Boca Raton, FL; Houslay and Milligan (1990) G-Proteins as Mediators of Cellular Signaling Processes Wiley and Sons, New York, NY; and Dohlman, et al. (1991) Ann. Rev. Biochem. 60:653-688. These hydrophobic

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domains are predicted to represent transmembrane spanning regions of the proteins. These GPCRs are found in a wide range of organisms and are typically involved in the transmission of signals to the interior of the cell, e.g., through interaction, e.g., with heterotrimeric G-proteins. They respond to a wide and diverse range of agents including lipid analogs, amino acid derivatives, small peptides, and other molecules.

The presumed transmembrane segments are typically 20-25 amino acids in length. Based upon models and data on bacteriorhodopsin, these regions are predicted to be a-helices and be oriented to form a ligand binding pocket. See, e.g., Findley, et al. (1990) Trends Pharmacol. Sci. 11:492-499. Other data suggest that the amino termini of the proteins are extracellular, and the carboxy termini are intracellular. See, e.g., Lodish, et al. (1995) Molecular Cell Biology 3d ed., Scientific American, New York; and Watson and Arkinstall (1994) The G-Protein Linked Receptor FactsBook Academic Press, San Diego, CÁ. Phosphorylation cascades have been implicated in the signal transduction pathway of these receptors.

Although the full spectrum of biological activities mediated by these 7 transmembrane receptors has not been fully determined, chemoattractant effects are recognized. Chemokine receptors are notable members of the GPCR family. See, e.g., Samson, et al. (1996) Biochemistry 35:3362-3367; and Rapport, et al. (1996) I. Leukocyte Biology 59:18-23. The best known biological functions of these chemokine molecules relate to chemoattraction of leukocytes. However, new chemokines and receptors are being discovered, and their biological effects on the various cells responsible for immunological responses are topics of continued study.

Many factors have been identified which influence the differentiation process of precursor cells, or regulate the physiology or migration properties of specific cell types. These observations indicate that other factors exist whose functions in immune function were heretofore unrecognized. These factors provide for biological activities whose spectra of effects may be distinct from known differentiation or activation factors. The absence of knowledge about the structural, biological, and physiological

properties of the regulatory factors which regulate cell physiology in vivo prevents the modulation of the effects of such factors.

In addition, other factors exist whose functions in hematopoiesis, neural function, immune development, and leukocyte trafficking were heretofore unrecognized. These receptors mediate biological activities whose spectra of effects are distinct from known differentiation, activation, or other signaling factors. The absence of knowledge about the structural, biological, and physiological properties of the receptors which regulate cell physiology, development, or function prevents the modification of the effects of such factors.

Thus, medical conditions where regulation of the development or physiology of relevant cells is required remain unmanageable.

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SUMMARY OF THE INVENTION

The present invention is based, in part, upon the discovery of new genes encoding various chemokines, e.g., those designated IBICK, which encodes primate CXC chemokines; ILINCK, which encodes primate CXC chemokines; CXC-143, which encodes rodent CXC chemokines; MCP243, which encodes a mouse chemokine; or 7 transmembrane receptors, e.g., those designated R277, which encode primate receptors; HST01.1, which encode rodent receptors; and 941D12, which encode rodent receptors. Each GPCR gene encodes a polypeptide exhibiting structural and/or sequence homology to 7 transmembrane receptors. Such receptors are typically G-protein coupled (or linked) receptors (GPCR or GPLR), though a ligand for each has not yet been identified.

The invention also provides mutations (muteins) of the respective natural sequences, fusion proteins, chemical mimetics, antibodies, and other structural or functional analogs. It is also directed to isolated nucleic acids, e.g., genes encoding respective proteins of the invention. Various uses of these different protein, antibody, or nucleic acid compositions are also provided.

The present invention provides a composition selected from the group of: a substantially pure antigenic polypeptide comprising sequence from an IBICK; an ILINCK; a CXC-143; an MCP243; an R277; an HST01.1; or a 941D12; a binding composition comprising an antigen binding portion of an antibody specific for binding to such an antigenic polypeptide; a nucleic acid encoding such an antigenic polypeptide; and a fusion protein comprising at least two non-overlapping segments of at least 10 amino acids of such an antigenic polypeptide.

In certain embodiments of the antigenic polypeptide, it is from a warm blooded animal, e.g., a rodent or primate; it comprises a sequence of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26; it exhibits a post-translational modification pattern distinct from a natural form of said polypeptide; it is detectably labeled; or it is made by expression of a recombinant nucleic acid. In other embodiments, a sterile form is provided, including, e.g., composition comprising the polypeptide and an acceptable carrier. A detection kit comprising

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a compartment or container holding such an antigenic polypeptide is also provided.

In other binding composition forms, e.g., antibody embodiments, the polypeptide is a mouse or human protein; the antibody is raised against a peptide sequence of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26; the antibody is a monoclonal antibody; the binding composition is fused to a heterologous protein, or is detectably labeled. An alternative embodiment is a binding compound comprising an antigen binding fragment of the antibody described. Also provided is a detection kit comprising such a binding compound. With the antibodies are provided methods of purifying a polypeptide using the binding compound or antibody to specifically separate the polypeptides from others, or for detection, e.g., immunohistochemistry or immunoprecipitation.

Nucleic acid embodiments are provided, e.g., where the nucleic acid is in an expression vector and: encodes a polypeptide from a mouse or human; encodes a mature protein of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26; or comprises a deoxyribonucleic acid nucleotide. The invention also provides a kit with such nucleic acids, e.g., which include PCR primers for amplifying such sequences.

With nucleic acids are provided fusion proteins, comprising: a sequence of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26; and/or sequence of another chemokine or 7 transmembrane receptor, as appropriate. Also provided is a cell comprising a recombinant nucleic acid, as described, and methods of producing a polypeptide comprising expressing the nucleic acid in an expression system.

Other embodiments include methods of modulating physiology or development of a cell, with a step of contacting that cell with a composition comprising an agonist or antagonist of the chemokine or receptor. Ordinarily, the cell is a neuron, macrophage, or lymphocyte. Various physiological effects to be modulated include a cellular calcium flux, a chemoattractant response, cellular morphology modification responses, phosphoinositide lipid turnover, or an antiviral response.

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

I. General

The present invention provides DNA sequences encoding various mammalian proteins, including chemokines, or which exhibit structural properties characteristic of a 7 transmembrane receptor. See, e.g., Ruffolo and Hollinger (eds. 1995) G-Protein Coupled Transmembrane Signaling Mechanisms CRC Press, Boca Raton, FL; Watson and Arkinstall (1994) The G-Protein Linked Receptor FactsBook Academic Press, San Diego, CA; Peroutka (ed. 1994) G Protein-Coupled Receptors CRC Press, Boca Raton, FL; Houslay and Milligan (1990) G-Proteins as Mediators of Cellular Signaling Processes Wiley and Sons, New York, NY. Certain human and mouse embodiments are described herein.

Among the many types of ligands which mediate biology via these receptors are chemokines and certain proteases. Chemokines play an important role in immune and inflammatory responses by inducing migration and adhesion of leukocytes. See, e.g., Schall (1991) Cytokine 3:165-183; and Thomson (ed.) The Cytokine Handbook Academic Press, NY. Chemokines are secreted by activated leukocytes and act as a chemoattractant for a variety of cells which are involved in inflammation. Besides chemoattractant properties, chemokines have been shown to induce other biological responses, e.g., modulation of second messenger levels such as Ca++; inositol phosphate pool changes (see, e.g., Berridge (1993) Nature 361:315-325 or Billah and Anthes (1990) Biochem. J. 269:281-291); cellular morphology modification responses; phosphoinositide lipid turnover; possible antiviral responses; and others. Thus, the chemokines provided herein may, alone or in combination with other therapeutic reagents, have advantageous combination effects.

Moreover, there are reasons to suggest that chemokines may have effects on other cell types, e.g., attraction or activation of monocytes, dendritic cells, T cells, eosinophils, and/or perhaps on basophils and/or neutrophils. They may also have chemoattractive effects on various neural cells including, e.g., dorsal root ganglia

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neurons in the peripheral nervous system and/or central nervous system neurons.

G-protein coupled receptors, e.g., chemokine receptors, are important in the signal transduction mechanisms mediated by their ligands. They are useful markers for distinguishing cell populations, and have been implicated as specific receptors for retroviral infections.

The chemokine superfamily was classically divided into two groups exhibiting characteristic structural motifs, the Cys-X-Cys (C-X-C) and Cys-Cys (C-C) families. These were distinguished on the basis of a single amino acid insertion between the NH-proximal pair of cysteine residues and sequence similarity. Typically, the C-X-C chemokines, i.e., IL-8 and MGSA/Gro-a act on neutrophils but not on monocytes, whereas the C-C chemokines, i.e., MIP-1a and RANTES, are potent chemoattractants for monocytes lymphocytes but not neutrophils. See, e.g., Miller, et al. (1992) Crit. Rev. Immunol. 12:17-46. A recently isolated chemokine, lymphotactin, does not belong to either group and may constitute a first member of a third chemokine family, the C family. Lymphotactin does not have a characteristic CC or CXC motif, and acts on lymphocytes but not neutrophils and monocytes. See, e.g., Kelner et al. (1994) Science 266:1395-1399. This chemokine defines a new C-C chemokine family. Even more recently, another chemokine exhibiting a CX3C motif has been identified, which establishes a fourth structural class.

The present invention provides additional chemokine reagents, e.g., nucleic acids, proteins and peptides, antibodies, etc., related to the newly discovered chemokines designated primate IBICK; primate ILINCK; rodent CXC-143; or rodent MCP243.

In other embodiments, the invention provides genes encoding novel G-protein coupled receptors, designated primate R277, rodent HST01.1, and rodent 941D12. Their ligands have not yet specifically been identified. However, the receptors exhibit structural features typical of known 7 transmembrane spanning receptors, which receptors include chemokine receptors. The receptors may exhibit properties of binding many different cytokines

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at varying specificities (shared or promiscuous binding specificity) or may exhibit high affinity for one (specific) or a subset (shared) of chemokines. Alternatively, the ligands may be other molecules, including molecules such as epinephrine, serotonin, or glucagon.

The described chemokines or receptors should be important for mediating various aspects of cellular, organ, tissue, or organismal physiology or development.

II. Purified Chemokines; Receptors

Nucleotide and derived amino acid sequences of a human embodiment of a primate CXC chemokine, designated IBICK are shown in SEQ ID NO: 1 and 2. The term "IBICK" will encompass other primate counterparts. The gene encodes a novel protein exhibiting structure and motifs characteristic of a chemokine. The predicted signal cleavage site is around the gly(-1)-phe1 peptide bond. Complementary nucleic acid sequences may be used for many purposes, e.g., in a PCR primer pair or as a mutagenesis primer. Fragments of the nucleotide sequence may be used as hybridization probes, or PCR primers, or to encode antigenic peptides. Fragments of the polypeptide will be useful as antigenic peptides. Likewise for the other genes. The closest reported chemokines to the IBICK sequences are the MIG and IP10 chemokines, both of which are IFN-γ regulated. See, e.g., Faubert (1993) Biochem. Biophys. Res. Commun. 192:223-230; and Luster, et al. (1985) Nature 315:672-676.

Nucleotide and derived amino acid sequences of a novel primate CC chemokine, e.g., from human, designated ILINCK are shown in SEQ ID NO: 3 and 4. The term "ILINCK" as used in this filing will encompass other primate counterparts. The predicted signal cleavage site is around the ser(-1)-gln1 peptide bond. Two different messages have been detected which encode the chemokine, and the larger one, a 1.5 kB message, is upregulated by IL-10. This is an unusual property of chemokine messages, which implies that the chemokine has a role in anti-inflammatory responses.

Partial nucleotide and derived amino acid sequences of a novel rodent CXC chemokine, e.g., from mouse, designated CXC-143, are shown in SEQ ID NO: 5, 6, 7, 8, 9 and 10. The term "CXC-143"

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will typically encompass rodent counterparts. Sequence analysis shows closest sequence homology to the IP10, MIG, and IBICK chemokines, described above. It may well be that the chemokine will be similarly regulated, e.g., by IFN-γ.

Nucleotide and derived amino acid sequences of a novel rodent chemokine, e.g., from mouse, designated MCP243 are shown in SEQ ID NO: 11, 12, 13 and 14. The term may encompass other rodent counterparts. Cys residues 14 and 30 correspond to conserved chemokine Cys3 and Cys4. Sequence analysis shows closest sequence homology to other chemokines.

Nucleotide and derived amino acid sequences of a novel rodent GPCR, e.g., from mouse, designated R277, are shown in SEQ ID NO: 15, 16, 17 and 18. The term "R277" may encompass other primate counterparts. Note that nucleotide 447 is designated C, but may be C or T; nucleotides 489 and 640 are each designated C, but may be A, C, G, or T; and nucleotides 480-510 may contain various sequence errors, but which will retain reading frame.. Sequence analysis shows closest sequence homology to a human GPCR designated TDAG8.

Partial nucleotide and derived amino acid sequences of a novel rodent GPCR, e.g., from mouse, designated HST01.1, are shown in SEQ ID NO: 19 and 20. The term "R277" may encompass other primate counterparts. Note that nucleotide 447 is designated C, but may be C or T; nucleotides 489 and 640 are each designated C, but may be A, C, G, or T; and nucleotides 480-510 may contain various sequence errors, but which will retain reading frame. The sequence is supplemented with more complete sequence in SEQ ID NO: 21 and 22. A DRY box motif runs from about asp147 to ala155; transmembrane segments run from about ala57 to leu78; phe90 to val110; val125 to phe146; val167 to leu189; phe223 to val243; leu255 to leu279; and val301 to val322. Sequence analysis shows sequence homology to various GPCR family members.

Partial nucleotide and derived amino acid sequences of a novel rodent GPCR, e.g., from mouse, designated 941D12, are shown in SEQ ID NO: 23 and 24. The term "941D12" may encompass other rodent counterparts. The nucleotides at positions 169, 178, 217, 287,

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290, 382, 386, 395, 411, 484, 512, 515, 517, and 521 are each indicated as C, but may be A, C, G, or T. A complete rodent 941D12 is provided in SEQ ID NO: 25 and 26. Nucleotide 942 is designated C, but may be C or T; nucleotides 1412 and 1422 each designated C, but may be A, C, G, or T. Sequence analysis shows sequence homology to various GPCR family members.

Certain general descriptions of physical properties of polypeptides, nucleic acids, and antibodies, where directed to one embodiment clearly are usually applicable to other chemokines or receptors described herein.

These amino acid sequences, provided amino to carboxy, are important in providing sequence information on the chemokine ligand or receptor, allowing for distinguishing the protein from other proteins, particularly naturally occurring versions. Moreover, the sequences allow preparation of peptides to generate antibodies to recognize and distinguish such segments, and allow preparation of oligonucleotide probes, both of which are strategies for isolation, e.g., cloning, of genes encoding such sequences, or related sequences, e.g., natural polymorphic or other variants, including fusion proteins. Similarities of the chemokines have been observed with other cytokines. See, e.g., Bosenberg, et al. (1992) Cell 71:1157-1165; Huang, et. al. (1992) Molecular Biology of the Cell 3:349-362; and Pandiella, et al. (1992) I. Biol. Chem. 267:24028-24033. Likewise for the GPC receptors.

As used herein, the term "IBICK" shall encompass, when used in a protein context, a protein having mature amino acid sequence, as shown in SEQ ID NO: 2. The invention also embraces a polypeptide comprising a significant fragment of such protein. The invention also encompasses a polypeptide which is a primate species counterpart, e.g., which exhibits similar sequence, and is more homologous in natural encoding sequence than other genes from a primate species. Typically, such chemokine will also interact with its specific binding components, e.g., receptor, or antibodies which bind to it. These binding components, e.g., antibodies, typically bind to the chemokine with high affinity, e.g., at least about 100 nM, usually better than about 30 nM, preferably better than about 10 nM, and

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more preferably at better than about 3 nM. Similar concepts apply to the primate embodiments for the chemokine ILINCK and the GPCR R277. In contrast, rodent embodiments for the chemokines CXC-143 and MCP243, and the GPCRs HST01.1 and 941D12 encompass other rodent species counterparts.

The term "polypeptide" as used herein includes a significant fragment or segment, and encompasses a stretch of amino acid residues of at least about 8 amino acids, generally at least 10 amino acids, more generally at least 12 amino acids, often at least 14 amino acids, more often at least 16 amino acids, typically at least 18 amino acids, more typically at least 20 amino acids, usually at least 22 amino acids, more usually at least 24 amino acids, preferably at least 26 amino acids, more preferably at least 28 amino acids, and, in particularly preferred embodiments, at least about 30 or more amino acids, e.g., about 35, 40, 45, 50, 60, 75, 80, 100, 120, etc. Similar proteins will likely comprise a plurality of such segments. Such fragments may have ends which begin and/or end at virtually all positions, e.g., beginning at residues 1, 2, 3, etc., and ending at, e.g., 69, 68, 67, 66, etc., in all combinatorial pairs in the coding segment. Particularly interesting peptides have ends corresponding to structural domain boundaries, e.g., intracellular or extracellular loops of the receptor embodiments. Such peptides will typically be immunogenic peptides, or may be concatenated to generate larger polypeptides. Short peptides may be attached or coupled to a larger carrier.

with specificity to the respective chemokine or receptor, e.g., in a ligand-receptor type fashion or an antibody-antigen interaction. These compositions may be compounds, e.g., proteins, which specifically associate with the chemokine or receptor, including natural physiologically relevant protein-protein interactions, either covalent or non-covalent. The binding composition may be a polymer, or another chemical reagent. No implication as to whether the chemokine presents a concave or convex shape in its ligand-receptor interaction is necessarily represented, other than the interaction exhibit similar specificity, e.g., specific affinity. A

functional analog may be a ligand with structural modifications, or

The term "binding composition" refers to molecules that bind

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may be a wholly unrelated molecule, e.g., which has a molecular shape which interacts with the appropriate ligand binding determinants. The ligands may serve as agonists or antagonists of a physiological or natural receptor, see, e.g., Goodman, et al. (eds.) (1990) Goodman & Gilman's: The Pharmacological Bases of Therapeutics (8th ed.), Pergamon Press. The term expressly includes antibodies, polyclonal or monoclonal, which specifically bind to the respective antigen.

Substantially pure means that the protein is free from other contaminating proteins, nucleic acids, and/or other biologicals typically derived from the original source organism. Purity may be assayed by standard methods, and will ordinarily be at least about 40% pure, more ordinarily at least about 50% pure, generally at least about 60% pure, more generally at least about 70% pure, often at least about 75% pure, more often at least about 80% pure, typically at least about 85% pure, more typically at least about 90% pure, preferably at least about 95% pure, more preferably at least about 98% pure, and in most preferred embodiments, at least 99% pure. Analyses will typically be by weight, but may be by molar amounts.

Solubility of a polypeptide or fragment depends upon the environment and the polypeptide. Many parameters affect polypeptide solubility, including temperature, electrolyte environment, size and molecular characteristics of the polypeptide, and nature of the solvent. Typically, the temperature at which the polypeptide is used ranges from about 4° C to about 65° C. Usually the temperature at use is greater than about 18° C and more usually greater than about 22° C. For diagnostic purposes, the temperature will usually be about room temperature or warmer, but less than the denaturation temperature of components in the assay. therapeutic purposes, the temperature will usually be body temperature, typically about 37° C for humans, though under certain situations the temperature may be raised or lowered in situ or in vitro.

The electrolytes will usually approximate in situ physiological conditions, but may be modified to higher or lower ionic strength where advantageous. The actual ions may be modified, e.g., to

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conform to standard buffers used in physiological or analytical contexts.

The size and structure of the polypeptide should generally be in a substantially stable state, and usually not in a denatured state, though in certain circumstances denatured protein will be important. The polypeptide may be associated with other polypeptides in a quaternary structure, e.g., to confer solubility, or associated with lipids or detergents in a manner which approximates natural lipid bilayer interactions.

The solvent will usually be a biologically compatible buffer, of a type used for preservation of biological activities, and will usually approximate a physiological solvent. Usually the solvent will have a neutral pH, typically at least about 5, preferably at least 6, and typically less than 10, preferably less than 9, and more preferably about 7.5. On some occasions, a detergent will be added, typically a mild non-denaturing one, e.g., CHS (cholesteryl hemisuccinate) or CHAPS (3-([3-cholamido-propyl]dimethylammonio)-1-propane sulfonate), or a low enough concentration as to avoid significant disruption of structural or physiological properties of the protein.

Solubility is reflected by sedimentation measured in Svedberg units, which are a measure of the sedimentation velocity of a molecule under particular conditions. The determination of the sedimentation velocity was classically performed in an analytical ultracentrifuge, but is typically now performed in a standard ultracentrifuge. See, Freifelder (1982) Physical Biochemistry (2d ed.), W.H. Freeman; and Cantor and Schimmel (1980) Biophysical Chemistry, parts 1-3, W.H. Freeman & Co., San Francisco. As a crude determination, a sample containing a putatively soluble polypeptide is spun in a standard full sized ultracentrifuge at about 50K rpm for about 10 minutes, and soluble molecules will remain in the supernatant. A soluble particle or polypeptide will typically be less than about 30S, more typically less than about 15S, usually less than about 10S, more usually less than about 6S, and, in particularembodiments, preferably less than about 4S, and more preferably less than about 3S.

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III. Physical Variants

This invention also encompasses proteins or peptides having substantial amino acid sequence homology with the amino acid sequence of each respective receptor. The variants include species or polymorphic variants.

Amino acid sequence homology, or sequence identity, is determined by optimizing residue matches, if necessary, by introducing gaps as required. This changes when considering conservative substitutions as matches. Conservative substitutions typically include substitutions within the following groups: glycine, alanine; valine, isoleucine, leucine; aspartic acid, glutamic acid; asparagine, glutamine; serine, threonine; lysine, arginine; and phenylalanine, tyrosine. Homologous amino acid sequences are typically intended to include natural allelic and interspecies variations in each respective protein sequence. Typical homologous proteins or peptides will have from 25-100% homology (if gaps can be introduced), to 50-100% homology (if conservative substitutions are included) with the amino acid sequence of the appropriate chemokine or receptor. Homology measures will be at least about 35%, generally at least 40%, more generally at least 45%, often at least 50%, more often at least 55%, typically at least 60%, more typically at least 65%, usually at least 70%, more usually at least 75%, preferably at least 80%, and more preferably at least 80%, and in particularly preferred embodiments, at least 85% or more. See also Needleham, et al. (1970) J. Mol. Biol. 48:443-453; Sankoff, et al. (1983) Chapter One in Time Warps, String Edits, and Macromolecules: The Theory and Practice of Sequence Comparison Addison-Wesley, Reading, MA; and software packages from IntelliGenetics, Mountain View, CA; and the University of Wisconsin Genetics Computer Group, Madison, WI.

Each of the isolated chemokine or GPC receptor DNAs can be readily modified by nucleotide substitutions, nucleotide deletions, nucleotide insertions, and inversions of nucleotide stretches. These modifications may result in novel DNA sequences which encode these antigens, their derivatives, or proteins having similar physiological, immunogenic, or antigenic activity. These modified

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sequences can be used to produce mutant antigens or to enhance expression, or to introduce convenient enzyme recognition sites into the nucleotide sequence without significantly affecting the encoded protein sequence. Enhanced expression may involve amplification, increased transcription, increased translation, and other mechanisms. Such mutant receptor derivatives include predetermined or site-specific mutations of the respective protein or "Mutant chemokine" encompasses a polypeptide otherwise falling within the homology definition of the chemokine as set forth above, but having an amino acid sequence which differs from that of the chemokine as found in nature, whether by way of deletion, substitution, or insertion. Likewise for the GPCRs. These include amino acid residue substitution levels from none, one, two, three, five, seven, ten, twelve, fifteen, etc. In particular, "site specific mutant" generally includes proteins having significant homology with a protein having sequences of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26, and as sharing various biological activities, e.g., antigenic or immunogenic, with those sequences, and in preferred embodiments contain most of the disclosed sequences, particularly those found in various groups of animals. As stated before, it is emphasized that descriptions are generally meant to encompass the various chemokine or receptor proteins from other members of related groups, not limited to the mouse or human embodiments specifically discussed.

Although site specific mutation sites are often predetermined, mutants need not be site specific. Chemokine or receptor mutagenesis can be conducted by making amino acid insertions or deletions. Substitutions, deletions, insertions, or combinations may be generated to arrive at a final construct. Insertions include amino-or carboxy- terminal fusions. Random mutagenesis can be conducted at a target codon and the expressed mutants can then be screened for the desired activity. Methods for making substitution mutations at predetermined sites in DNA having a known sequence are well known in the art, e.g., by M13 primer mutagenesis or polymerase chain reaction (PCR) techniques. See also Sambrook, et al. (1989) and Ausubel, et al. (1987 and Supplements). Many

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structural features are known about the chemokines and GPCRs which allow determination of whether specific residues are embedded into the core of the secondary or tertiary structures, or whether the residues will have relatively little effect on protein folding. Preferred positions for mutagenesis are those which do not prevent functional folding of the resulting protein.

The mutations in the DNA normally should not place coding sequences out of reading frames and preferably will not create complementary regions that could hybridize to produce secondary mRNA structure such as loops or hairpins. But certain situations exist where such problems are compensated. See, e.g., Gesteland and Atkins (1996) Ann. Rev. Biochem. 65:741-768.

The present invention also provides recombinant proteins, e.g., heterologous fusion proteins using segments from these proteins, or antibodies. A heterologous fusion protein is a fusion of proteins or segments which are naturally not normally fused in the same manner. Thus, the fusion product of an immunoglobulin with a receptor polypeptide is a continuous protein molecule having sequences fused in a typical peptide linkage, typically made as a single translation product and exhibiting properties derived from each source peptide. A similar chimeric concept applies to heterologous nucleic acid sequences.

In addition, new constructs may be made from combining similar functional or structural domains from other proteins. For example, ligand-binding or other segments may be "swapped" between different new fusion polypeptides or fragments. See, e.g., Cunningham, et al. (1989) Science 243:1330-1336; and O'Dowd, et al. (1988) <u>J. Biol. Chem.</u> 263:15985-15992. Thus, new chimeric polypeptides exhibiting new combinations of specificities will result from the functional linkage of ligand-binding specificities and other functional domains. Such may be chimeric molecules with mixing or matching of the various structural segments, e.g., the β -sheet or α helix structural domains for the chemokine, or receptor segments corresponding to each of the transmembrane segments (TM1-TM7),

35 or the intracellular (cytosolic, C1-C4) or extracellular (E1-E4) loops from the various receptor types. The C3 loop is particularly important.

The phosphoramidite method described by Beaucage and Carruthers (1981) <u>Tetra. Letts.</u> 22:1859-1862, will produce suitable synthetic DNA fragments. A double stranded fragment will often be obtained either by synthesizing the complementary strand and annealing the strand together under appropriate conditions or by adding the complementary strand using DNA polymerase with an appropriate primer sequence, e.g., PCR techniques.

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IV. Functional Variants

The blocking of physiological response to various embodiments of these chemokines or GPCRs may result from the inhibition of binding of the ligand to its receptor, likely through competitive inhibition. Thus, in vitro assays of the present invention will often use isolated protein, membranes from cells expressing a recombinant membrane associated receptor, e.g., ligand binding segments, or fragments attached to solid phase substrates. These assays will also allow for the diagnostic determination of the effects of either binding segment mutations and modifications, or ligand mutations and modifications, e.g., ligand analogs.

This invention also contemplates the use of competitive drug screening assays, e.g., where neutralizing binding compositions, e.g., antibodies, to antigen or receptor fragments compete with a test compound for binding to the protein. In this manner, the antibodies can be used to detect the presence of polypeptides which share one or more antigenic binding sites of the ligand and can also be used to occupy binding sites on the protein that might otherwise interact with a receptor.

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Additionally, neutralizing antibodies against a specific chemokine embodiment and soluble fragments of the chemokine which contain a high affinity receptor binding site, can be used to inhibit chemokine activity in tissues, e.g., tissues experiencing abnormal physiology.

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"Derivatives" of chemokine antigens include amino acid sequence mutants, glycosylation variants, and covalent or aggregate

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conjugates with other chemical moieties. Covalent derivatives can be prepared by linkage of functionalities to groups which are found in chemokine amino acid side chains or at the N- or C- termini, by means which are well known in the art. These derivatives can include, without limitation, aliphatic esters or amides of the carboxyl terminus, or of residues containing carboxyl side chains, O-acyl derivatives of hydroxyl group-containing residues, and N-acyl derivatives of the amino terminal amino acid or amino-group containing residues, e.g., lysine or arginine. Acyl groups are selected from the group of alkyl-moieties including C3 to C18 normal alkyl, thereby forming alkanoyl aroyl species. Covalent attachment to carrier proteins may be important when immunogenic moieties are haptens.

In particular, glycosylation alterations are included, e.g., made by modifying the glycosylation patterns of a polypeptide during its synthesis and processing, or in further processing steps. Particularly preferred means for accomplishing this are by exposing the polypeptide to glycosylating enzymes derived from cells which normally provide such processing, e.g., mammalian glycosylation enzymes. Deglycosylation enzymes are also contemplated. Also embraced are versions of the same primary amino acid sequence which have other minor modifications, including phosphorylated amino acid residues, e.g., phosphotyrosine, phosphoserine, or phosphothreonine, or nucleoside or nucleotide derivatives, e.g., guanyl derivatized.

A major group of derivatives are covalent conjugates of the respective chemokine or receptor or fragments thereof with other proteins or polypeptides. These derivatives can be synthesized in recombinant culture such as N- or C-terminal fusions or by the use of agents known in the art for their usefulness in cross-linking proteins through reactive side groups. Preferred chemokine derivatization sites with cross-linking agents are at free amino groups, carbohydrate moieties, and cysteine residues.

Fusion polypeptides between these chemokines or receptors and other homologous or heterologous proteins, e.g., other chemokines or receptors, are also provided. Many growth factors

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and cytokines are homodimeric entities, and a repeat construct may have various advantages, including lessened susceptibility to proteolytic cleavage. Moreover, many cytokine receptors require dimerization to transduce a signal, and various dimeric ligands or domain repeats can be desirable. Homologous polypeptides may be fusions between different surface markers, resulting in, e.g., a hybrid protein exhibiting receptor binding specificity. heterologous fusions may be constructed which would exhibit a combination of properties or activities of the derivative proteins. Typical examples are fusions of a reporter polypeptide, e.g., luciferase, with a segment or domain of a ligand, e.g., a receptorbinding segment, so that the presence or location of the fused ligand, or a binding composition, may be easily determined. See, e.g., Dull, et al., U.S. Patent No. 4,859,609. Other gene fusion partners include bacterial B-galactosidase, trpE, Protein A, B-lactamase, alpha amylase, alcohol dehydrogenase, a FLAG fusion, and yeast alpha mating factor. See, e.g., Godowski, et al. (1988) Science 241:812-816.

The phosphoramidite method described by Beaucage and Carruthers (1981) <u>Tetra. Letts.</u> 22:1859-1862, will produce suitable synthetic DNA fragments. A double stranded fragment will often be obtained either by synthesizing the complementary strand and annealing the strand together under appropriate conditions or by adding the complementary strand using DNA polymerase with an appropriate primer sequence.

Such polypeptides may also have amino acid residues which have been chemically modified by phosphorylation, guanylation, sulfonation, biotinylation, or the addition or removal of other moieties, particularly those which have molecular shapes similar to phosphate or guanyl groups. In some embodiments, the modifications will be useful labeling reagents, or serve as purification targets, e.g., affinity tags as FLAG.

Fusion proteins will typically be made by either recombinant nucleic acid methods or by synthetic polypeptide methods. Techniques for nucleic acid manipulation and expression are described generally, for example, in Sambrook, et al. (1989) Molecular Cloning: A Laboratory Manual (2d ed.), Vols. 1-3, Cold Spring

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Harbor Laboratory. Techniques for synthesis of polypeptides are described, for example, in Merrifield (1963) <u>I. Amer. Chem. Soc.</u> 85:2149-2156; Merrifield (1986) <u>Science</u> 232: 341-347; and Atherton, et al. (1989) <u>Solid Phase Peptide Synthesis: A Practical Approach</u>, IRL Press, Oxford; and chemical ligation, e.g., Dawson, et al. (1994) <u>Science</u> 266:776-779, a method of linking long synthetic peptides by a peptide bond.

This invention also contemplates the use of derivatives of these chemokines or receptors other than variations in amino acid sequence or glycosylation. Such derivatives may involve covalent or aggregative association with chemical moieties. derivatives generally include: (1) salts, (2) side chain and terminal residue covalent modifications, and (3) adsorption complexes, for example with cell membranes. Such covalent or aggregative derivatives are useful as immunogens, as reagents immunoassays, or in purification methods such as for affinity purification of ligands or other binding ligands. For example, a chemokine antigen can be immobilized by covalent bonding to a solid support such as cyanogen bromide-activated Sepharose, by methods which are well known in the art, or adsorbed onto polyolefin surfaces, with or without glutaraldehyde cross-linking, for use in the assay or purification of anti-chemokine antibodies or its receptor. These chemokines can also be labeled with a detectable group, for example radioiodinated by the chloramine T procedure, covalently bound to rare earth chelates, or conjugated to a fluorescent moiety for use in diagnostic assays. Purification of chemokine, receptor, or binding compositions may be effected by immobilized antibodies or receptor.

Other modifications may be introduced with the goal of modifying the therapeutic pharmacokinetics or pharmacodynamics of a target chemokine. For example, certain means to minimize the size of the entity may improve its pharmacoaccessibility; other means to maximize size may affect pharmacodynamics. Similarly, changes in ligand binding kinetics or equilibrium of a receptor may be engineered.

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A solubilized chemokine or receptor or appropriate fragment of this invention can be used as an immunogen for the production of antisera or antibodies specific for the ligand, receptor, or fragments thereof. The purified proteins can be used to screen monoclonal antibodies chemokine-binding fragments prepared immunization with various forms of impure preparations containing the protein. In particular, antibody equivalents include antigen binding fragments of natural antibodies, e.g., Fv, Fab, or F(ab)2. Purified chemokines can also be used as a reagent to detect antibodies generated in response to the presence of elevated levels of the protein or cell fragments containing the protein, both of which may be diagnostic of an abnormal or specific physiological or disease Additionally, chemokine protein fragments, or their concatenates, may also serve as immunogens to produce binding compositions, e.g., antibodies of the present invention, as described immediately below. For example, this invention contemplates antibodies raised against certain amino acid sequences, e.g., in in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 and 26, or proteins In particular, this invention contemplates containing them. antibodies having binding affinity to or being raised against specific fragments, e.g., those which are predicted to lie on the outside surfaces of protein tertiary structure. Similar concepts apply to antibodies specific for receptors of the invention.

The present invention contemplates the isolation of additional closely related species variants. Southern and Northern blot analysis should establish that similar genetic entities exist in other related mammals, and establish the stringency of hybridization conditions to isolate such. It is likely that these chemokines and receptors are widespread in species variants, e.g., among the rodents and the primates.

The invention also provides means to isolate a group of related chemokines or receptors displaying both distinctness and similarities in structure, expression, and function. Elucidation of many of the physiological effects of the proteins will be greatly accelerated by the isolation and characterization of distinct species variants of the ligands. Related genes found, e.g., in various

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computer databases will also be useful, in many instances, for similar purposes with structurally related proteins. In particular, the present invention provides useful probes or search features for identifying additional homologous genetic entities in different species.

The isolated genes will allow transformation of cells lacking expression of a corresponding chemokine or receptor, e.g., either species types or cells which lack corresponding antigens and exhibit negative background activity. Expression of transformed genes will allow isolation of antigenically pure cell lines, with defined or single specie variants. This approach will allow for more sensitive detection and discrimination of the physiological effects of chemokine or receptor proteins. Subcellular fragments, e.g., cytoplasts or membrane fragments, can be isolated and used.

Dissection of critical structural elements which effect the various differentiation functions provided by ligands is possible using standard techniques of modern molecular biology, particularly in comparing members of the related class. See, e.g., the homolog-scanning mutagenesis technique described in Cunningham, et al. (1989) Science 243:1339-1336; and approaches used in O'Dowd, et al. (1988) J. Biol. Chem. 263:15985-15992; and Lechleiter, et al. (1990) EMBO J. 9:4381-4390.

In addition, various segments can be substituted between species variants to determine what structural features are important in both receptor binding affinity and specificity, as well as signal transduction. An array of different chemokine or receptor variants will be used to screen for variants exhibiting combined properties of interaction with different species variants.

Intracellular functions would probably involve segments of the receptor which are normally accessible to the cytosol. However, ligand internalization may occur under certain circumstances, and interaction between intracellular components and "extracellular" segments may occur. The specific segments of interaction of a particular chemokine with other intracellular components may be identified by mutagenesis or direct biochemical means, e.g., cross-linking or affinity methods. Structural analysis by crystallographic

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or other physical methods will also be applicable. Further investigation of the mechanism of signal transduction will include study of associated components which may be isolatable by affinity methods or by genetic means, e.g., complementation analysis of mutants.

Further study of the expression and control of the various chemokines or receptors will be pursued. The controlling elements associated with the proteins may exhibit differential developmental, tissue specific, or other expression patterns. Upstream or downstream genetic regions, e.g., control elements, are of interest. Differential splicing of message may lead to membrane bound forms, soluble forms, and modified versions of ligand.

Structural studies of the proteins will lead to design of new ligands or receptors, particularly analogs exhibiting agonist or antagonist properties on the receptor. This can be combined with previously described screening methods to isolate ligands exhibiting desired spectra of activities.

Expression in other cell types will often result in glycosylation differences in a particular chemokine or receptor. Various species variants may exhibit distinct functions based upon structural differences other than amino acid sequence. Differential modifications may be responsible for differential function, and elucidation of the effects are now made possible.

Thus, the present invention provides important reagents related to a physiological ligand-receptor interaction. Although the foregoing description has focused primarily upon the mouse and human embodiments of the chemokines or receptors specifically described, those of skill in the art will immediately recognize that the invention provides other counterparts, e.g., from related species, rodents or primates.

V. Antibodies

Antibodies can be raised to these chemokines or receptors, including species or polymorphic variants, and fragments thereof, both in their naturally occurring forms and in their recombinant forms. Additionally, antibodies can be raised to chemokines or

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receptors in either their active or inactive forms, or in their native or denatured forms. Anti-idiotypic antibodies are also contemplated.

Antibodies, including binding fragments and single chain versions, against predetermined fragments of the ligands can be raised by immunization of animals with concatemers or conjugates of the fragments with immunogenic proteins. Monoclonal antibodies are prepared from cells secreting the desired antibody. These antibodies can be screened for binding to normal or defective chemokines or receptors, or screened for agonistic or antagonistic activity. These monoclonal antibodies will usually bind with at least a KD of about 1 mM, more usually at least about 300 μ M, typically at least about 10 μ M, more typically at least about 30 μ M, preferably at least about 10 μ M, and more preferably at least about 3 μ M or better.

The antibodies, including antigen binding fragments, of this invention can have significant preparative, diagnostic, or therapeutic value. They can be useful to purify or label the desired antigen in a sample, or may be potent antagonists that bind to ligand and inhibit binding to receptor or inhibit the ability of a ligand to elicit a biological response. They also can be useful as non-neutralizing antibodies and can be coupled to, or as fusion proteins with, toxins or radionuclides so that when the antibody binds to antigen, a cell expressing it, e.g., on its surface via receptor, is killed. Further, these antibodies can be conjugated to drugs or other therapeutic agents, either directly or indirectly by means of a linker, and may effect drug targeting. Antibodies to receptors may be more easily used to block ligand binding and/or signal transduction.

The antibodies of this invention can also be useful in diagnostic or reagent purification applications. As capture or non-neutralizing antibodies, they can be screened for ability to bind to the chemokines or receptors without inhibiting ligand-receptor binding. As neutralizing antibodies, they can be useful in competitive binding assays. They will also be useful in detecting or quantifying chemokine or receptors, e.g., in immunoassays. They may be used as purification reagents in immunoaffinity columns or as immunohistochemistry reagents.

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Ligand or receptor fragments may be concatenated or joined to other materials, particularly polypeptides, as fused or covalently joined polypeptides to be used as immunogens. Short peptides will preferably be made as repeat structures to increase size. A ligand and its fragments may be fused or covalently linked to a variety of immunogens, such as keyhole limpet hemocyanin, bovine serum albumin, tetanus toxoid, etc. See Microbiology, Hoeber Medical Division, Harper and Row, 1969; Landsteiner (1962) Specificity of Serological Reactions, Dover Publications, New York, and Williams, et al. (1967) Methods in Immunology and Immunochemistry, Vol. 1, Academic Press, New York, for descriptions of methods of preparing polyclonal antisera. A typical method involves hyperimmunization of an animal with an antigen. The blood of the animal is then collected shortly after the repeated immunizations and the gamma globulin fraction is isolated.

In some instances, it is desirable to prepare monoclonal antibodies from various mammalian hosts, such as mice, rodents, primates, humans, etc. Description of techniques for preparing such monoclonal antibodies may be found in, e.g., Stites, et al. (eds.) Basic and Clinical Immunology (4th ed.), Lange Medical Publications, Los Altos, CA, and references cited therein; Harlow and Lane (1988) Antibodies: A Laboratory Manual, CSH Press; Goding (1986) Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York; and particularly in Kohler and Milstein (1975) in Nature 256:495-497, which discusses one method of generating monoclonal antibodies. Summarized briefly, this method involves injecting an animal with an immunogen. The animal is then sacrificed and cells taken, e.g., from its spleen, which are then fused with myeloma cells. The result is a hybrid cell or "hybridoma" that is capable of reproducing in vitro. The population of hybridomas is then screened to isolate individual clones, each of which secrete a single antibody species to the immunogen. In this manner, the individual antibody species obtained are the products immortalized and cloned single B cells from the immune animal generated in response to a specific site recognized on the

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immunogenic substance. Large amounts of antibody may be derived from ascites fluid from an animal.

Other suitable techniques involve in vitro exposure of lymphocytes to the antigenic polypeptides or alternatively to selection of libraries of antibodies in phage or similar vectors. See, Huse, et al. (1989) "Generation of a Large Combinatorial Library of the Immunoglobulin Repertoire in Phage Lambda," Science 246:1275-1281; and Ward, et al. (1989) Nature 341:544-546. polypeptides and antibodies of the present invention may be used with or without modification, including chimeric or humanized Frequently, the polypeptides and antibodies will be labeled by joining, either covalently or non-covalently, a substance which provides for a detectable signal. A wide variety of labels and conjugation techniques are known and are reported extensively in both the scientific and patent literature. Suitable labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent moieties, chemiluminescent moieties, magnetic particles, and the like. Patents, teaching the use of such labels include U.S. Patent Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241. Also, recombinant immunoglobulins may be produced, see Cabilly, U.S. Patent No. 4,816,567; and Queen et al. (1989) Proc. Nat'l. Acad. Sci. 86:10029-10033.

The antibodies of this invention can also be used for affinity chromatography in isolating the protein. Columns can be prepared where the antibodies are linked to a solid support, e.g., particles, such as agarose, Sephadex, or the like, where a cell lysate may be passed through the column, the column washed, followed by increasing concentrations of a mild denaturant, whereby the purified chemokine protein will be released.

The antibodies may also be used to screen expression libraries for particular expression products. Usually the antibodies used in such a procedure will be labeled with a moiety allowing easy detection of presence of antigen by antibody binding.

Antibodies raised against these chemokines or receptors will also be useful to raise anti-idiotypic antibodies. These will be useful

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in detecting or diagnosing various immunological conditions related to expression of the respective antigens.

VI. Nucleic Acids

The described peptide sequences and the related reagents are useful in isolating a DNA clone encoding these chemokines or receptors, e.g., from a natural source. Typically, it will be useful in isolating a gene from another individual, and similar procedures will be applied to isolate genes from related species, e.g., rodents or primates. Cross hybridization will allow isolation of ligand from other closely related species. A number of different approaches should be available to successfully isolate a suitable nucleic acid clone. Similar concepts apply to the receptor embodiments.

The purified protein or defined peptides are useful for generating antibodies by standard methods, as described above. Synthetic peptides or purified protein can be presented to an immune system to generate monoclonal or polyclonal antibodies. See, e.g., Coligan (1991) Current Protocols in Immunology Wiley/Greene; and Harlow and Lane (1989) Antibodies: A Laboratory Manual Cold Spring Harbor Press. Alternatively, a chemokine or receptor may be used as a specific binding reagent, and advantage can be taken of its specificity of binding, much like an antibody would be used. The chemokine receptors are typically 7 transmembrane proteins, which could be sensitive to appropriate interaction with lipid or membrane. The signal transduction typically is mediated through a G-protein, through interaction with a G-protein coupled receptor.

For example, the specific binding composition could be used for screening of an expression library made from a cell line which expresses a particular chemokine. The screening can be standard staining of surface expressed ligand, or by panning. Screening of intracellular expression can also be performed by various staining or immunofluorescence procedures. The binding compositions could be used to affinity purify or sort out cells expressing the ligand.

The peptide segments can also be used to predict appropriate oligonucleotides to screen a library, e.g., to isolate species variants.

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The genetic code can be used to select appropriate oligonucleotides useful as probes for screening. n combination with polymerase chain reaction (PCR) techniques, synthetic oligonucleotides will be useful in selecting correct clones from a library. Complementary sequences will also be used as probes or primers. Anchored vector or poly-A complementary PCR techniques or complementary DNA of other peptides may be useful.

This invention contemplates use of isolated DNA or fragments to encode a biologically active corresponding chemokine In addition, this invention covers isolated or polypeptide. recombinant DNA which encodes a biologically active protein or polypeptide which is capable of hybridizing under appropriate conditions with the DNA sequences described herein. biologically active protein or polypeptide can be an intact ligand. receptor, or fragment, and have an amino acid sequence as disclosed in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24 or 26. Further, this invention covers the use of isolated or recombinant DNA, or fragments thereof, which encode proteins which are homologous to a chemokine or receptor or which was isolated using such a cDNA encoding a chemokine or receptor as a probe. The isolated DNA can have the respective regulatory sequences in the 5' and 3' flanks, e.g., promoters, enhancers, poly-A addition signals, and others.

An "isolated" nucleic acid is a nucleic acid, e.g., an RNA, DNA, or a mixed polymer, which is substantially separated from other components which naturally accompany a native sequence, e.g., ribosomes, polymerases, and flanking genomic sequences from the originating species. The term embraces a nucleic acid sequence which has been removed from its naturally occurring environment, and includes recombinant or cloned DNA isolates and chemically synthesized analogs or analogs biologically synthesized by heterologous systems. A substantially pure molecule includes isolated forms of the molecule.

An isolated nucleic acid will generally be a homogeneous composition of molecules, but will, in some embodiments, contain minor heterogeneity. This heterogeneity is typically found at the .15

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polymer ends or portions not critical to a desired biological function or activity.

A "recombinant" nucleic acid is defined either by its method of production or its structure. In reference to its method of production, e.g., a product made by a process, the process is use of recombinant nucleic acid techniques, e.g., involving human intervention in the nucleotide sequence, typically selection or production. Alternatively, it can be a nucleic acid made by generating a sequence comprising fusion of two fragments which are not naturally contiguous to each other, but is meant to exclude products of nature, e.g., naturally occurring purified forms. Thus, for example, products made by transforming cells with any unnaturally occurring vector is encompassed, as are nucleic acids comprising sequence derived using a synthetic oligonucleotide process. Such is often done to replace a codon with a redundant codon encoding the same or a conservative amino acid, while typically introducing or removing a sequence recognition site. Alternatively, it is performed to join together nucleic acid segments of desired functions to generate a single genetic entity comprising a desired combination of functions not found in the commonly available natural forms. Restriction enzyme recognition sites are often the target of such artificial manipulations, but other site specific targets, e.g., promoters, DNA replication sites, regulation sequences, control sequences, or other useful features may be incorporated by design. A similar concept is intended for a recombinant, e.g., fusion, polypeptide. Specifically included are synthetic nucleic acids which, by genetic code redundancy, encode polypeptides similar to fragments of these antigens, and fusions of sequences from various different species variants.

A significant "fragment" in a nucleic acid context is a contiguous segment of at least about 17 nucleotides, generally at least about 20 nucleotides, more generally at least about 23 nucleotides, ordinarily at least about 26 nucleotides, more ordinarily at least about 29 nucleotides, often at least about 32 nucleotides, more often at least about 35 nucleotides, typically at least about 38 nucleotides, more typically at least about 41 nucleotides, usually at least about 44

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nucleotides, more usually at least about 47 nucleotides, preferably at least about 50 nucleotides, more preferably at least about 53 nucleotides, and in particularly preferred embodiments will be at least about 56 or more nucleotides, e.g., 60, 65, 75, 85, 100, 120, 150, 200, 250, 300, 400, etc. Such fragments may have ends which begin and/or end at virtually all positions, e.g., beginning at nucleotides 1, 2, 3, etc., and ending at, e.g., 300, 299, 298, 287, etc., in combinatorial pairs. Particularly interesting polynucleotides have ends corresponding to structural domain boundaries.

A DNA which codes for a particular chemokine or receptor protein or peptide will be very useful to identify genes, mRNA, and cDNA species which code for related or homologous ligands or receptors, as well as DNAs which code for homologous proteins from different species. There are likely homologs in closely related species, e.g., rodents or primates. Various chemokine proteins should be homologous and are encompassed herein, as would be receptors. However, proteins can readily be isolated under appropriate conditions using these sequences if they are sufficiently homologous. Typically, primate chemokines or receptors are of particular interest.

This invention further covers recombinant DNA molecules and fragments having a DNA sequence identical to or highly homologous to the isolated DNAs set forth herein. In particular, the sequences will often be operably linked to DNA segments which control transcription, translation, and DNA replication. Alternatively, recombinant clones derived from the genomic sequences, e.g., containing introns, will be useful for transgenic studies, including, e.g., transgenic cells and organisms, and for gene therapy. See, e.g., Goodnow (1992) "Transgenic Animals" in Roitt (ed.) Encyclopedia of Immunology Academic Press, San Diego, pp. 1502-1504; Travis (1992) Science 256:1392-1394; Kuhn, et al. (1991) Science 254:707-710; Capecchi (1989) Science 244:1288; Robertson (1987)(ed.) Teratocarcinomas and Embryonic Stem Cells: A Practical Approach IRL Press, Oxford; and Rosenberg (1992) J. Clinical Oncology 10:180-199.

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Homologous nucleic acid sequences, when compared, exhibit significant similarity, or identity. The standards for homology in nucleic acids are either measures for homology generally used in the art by sequence comparison or based upon hybridization conditions. The hybridization conditions are described in greater detail below.

Substantial homology in the nucleic acid comparison context means either that the segments, or their complementary strands, when compared, are identical when optimally aligned, with appropriate nucleotide insertions or deletions, in at least about 50% of the nucleotides, generally at least about 56%, more generally at least about 59%, ordinarily at least about 62%, more ordinarily at least about 65%, often at least about 68%, more often at least about 71%, typically at least about 74%, more typically at least about 77%, usually at least about 80%, more usually at least about 85%, preferably at least about 90%, more preferably at least about 95 to 98% or more, and in particular embodiments, as high at about 99% or more of the nucleotides. Alternatively, substantial homology exists when the segments will hybridize under selective hybridization conditions, to a strand, or its complement, typically using a sequence derived from SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23 or 25. Typically, selective hybridization will occur when there is at least about 55% homology over a stretch of at least about 30 nucleotides, preferably at least about 65% over a stretch of at least about 25 nucleotides, more preferably at least about 75%, and most preferably at least about 90% over about 20 nucleotides. See, Kanehisa (1984) Nuc. Acids Res. 12:203-213. The length of homology comparison, as described, may be over longer stretches, and in certain embodiments will be over a stretch of at least about 17 nucleotides, usually at least about 20 nucleotides, more usually at least about 24 nucleotides, typically at least about 28 nucleotides, more typically at least about 40 nucleotides, preferably at least about 50 nucleotides, and more preferably at least about 75 to 100 or more nucleotides. PCR primers will generally have high levels of matches over potentially shorter lengths.

Stringent conditions, in referring to homology in the hybridization context, will be stringent combined conditions of salt,

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temperature, organic solvents, and other parameters, typically those controlled in hybridization reactions. Stringent temperature conditions will usually include temperatures in excess of about 30° C, more usually in excess of about 37° C, typically in excess of about 45° C, more typically in excess of about 55° C, preferably in excess of about 65° C, and more preferably in excess of about 70° C. Stringent salt conditions will ordinarily be less than about 1000 mM, usually less than about 500 mM, more usually less than about 400 mM, typically less than about 300 mM, preferably less than about 200 mM, and more preferably less than about 150 mM, e.g., 20-50 mM. However, the combination of parameters is much more important than the measure of any single parameter. See, e.g., Wetmur and Davidson (1968) <u>I. Mol. Biol.</u> 31:349-370.

Corresponding chemokines or receptors from other closely related species can be cloned and isolated by cross-species hybridization. Alternatively, sequences from a sequence data base may be recognized as having similarity. Homology may be very low between distantly related species, and thus hybridization of relatively closely related species is advisable. Alternatively, preparation of an antibody preparation which exhibits less species specificity may be useful in expression cloning approaches. PCR approaches using segments of conserved sequences will also be used.

VII. Making Chemokines or Receptors; Mimetics

DNA which encodes each respective chemokine, receptor, or fragments thereof can be obtained by chemical synthesis, screening cDNA libraries, or by screening genomic libraries prepared from a wide variety of cell lines or tissue samples.

This DNA can be expressed in a wide variety of host cells for the synthesis of a full-length ligand or fragments which can in turn, for example, be used to generate polyclonal or monoclonal antibodies; for binding studies; for construction and expression of modified molecules; for expression cloning or purification; and for structure/function studies. Each antigen or its fragments can be expressed in host cells that are transformed or transfected with appropriate expression vectors. These molecules can be substantially

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purified to be free of protein or cellular contaminants, other than those derived from the recombinant host, and therefore are particularly useful in pharmaceutical compositions when combined with a pharmaceutically acceptable carrier and/or diluent. The antigens or antibodies, or portions thereof, may be expressed as fusions with other proteins.

Expression vectors are typically self-replicating DNA or RNA constructs containing the desired antigen gene or its fragments, usually operably linked to suitable genetic control elements that are recognized in a suitable host cell. These control elements are capable of effecting expression within a suitable host. The specific type of control elements necessary to effect expression will depend upon the eventual host cell used. Generally, the genetic control elements can include a prokaryotic promoter system or a eukaryotic promoter expression control system, and typically include a transcriptional promoter, an optional operator to control the onset of transcription, transcription enhancers to elevate the level of mRNA expression, a sequence that encodes a suitable ribosome binding site, and sequences that terminate transcription and translation. Expression vectors also usually contain an origin of replication that allows the vector to replicate independently of the host cell.

The vectors of this invention contain DNA which encode embodiments of a chemokine, receptor, or a fragment thereof, typically encoding a biologically active polypeptide. The DNA can be under the control of a viral promoter and can encode a selection marker. This invention further contemplates use of such expression vectors which are capable of expressing eukaryotic cDNA coding for each chemokine or receptor in a prokaryotic or eukaryotic host, where the vector is compatible with the host and where the eukaryotic cDNA coding for the protein is inserted into the vector such that growth of the host containing the vector expresses the cDNA in question. Usually, expression vectors are designed for stable replication in their host cells or for amplification to greatly increase the total number of copies of the desirable gene per cell. It is not always necessary to require that an expression vector replicate in a host cell, e.g., it is possible to effect transient expression of the

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ligand or its fragments in various hosts using vectors that do not contain a replication origin that is recognized by the host cell. It is also possible to use vectors that cause integration of a chemokine or receptor gene or its fragments into the host DNA by recombination, or to integrate a promoter which controls expression of an endogenous gene.

Vectors, as used herein, comprise plasmids, viruses, bacteriophage, integratable DNA fragments, and other vehicles, including those which enable the integration of DNA fragments into the genome of the host. Expression vectors are specialized vectors which contain genetic control elements that effect expression of operably linked genes. Plasmids are the most commonly used form of vector but many other forms of vectors which serve an equivalent function and which are, or become, known in the art are suitable for use herein. See, e.g., Pouwels, et al. (1985 and Supplements) Cloning Vectors: A Laboratory Manual, Elsevier, N.Y., and Rodriquez, et al. (1988)(eds.) Vectors: A Survey of Molecular Cloning Vectors and Their Uses, Buttersworth, Boston, MA.

Transformed cells include cells, preferably mammalian, that have been transformed or transfected with a chemokine or receptor gene containing vector constructed using recombinant DNA techniques. Transformed host cells usually express the ligand, receptor, or its fragments, but for purposes of cloning, amplifying, and manipulating its DNA, do not need to express the protein. This invention further contemplates culturing transformed cells in a nutrient medium, thus permitting the protein to accumulate in the culture. The protein can be recovered, from the culture or from the culture medium, or from cell membranes.

For purposes of this invention, DNA sequences are operably linked when they are functionally related to each other. For example, DNA for a presequence or secretory signal is operably linked to a polypeptide if it is expressed as a preprotein or participates in directing the polypeptide to the cell membrane or in secretion of the polypeptide. A promoter is operably linked to a coding sequence if it controls the transcription of the polypeptide; a ribosome binding site is operably linked to a coding sequence if it is

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positioned to permit translation. Usually, operably linked means contiguous and in reading frame, however, certain genetic elements such as repressor genes are not contiguously linked but still bind to operator sequences that in turn control expression.

Suitable host cells include prokaryotes, lower eukaryotes, and higher eukaryotes. Prokaryotes include both gram negative and gram positive organisms, e.g., E. coli and B. subtilis. Lower eukaryotes include yeasts, e.g., S. cerevisiae and Pichia, and species of the genus Dictyostelium. Higher eukaryotes include established tissue culture cell lines from animal cells, both of non-mammalian origin, e.g., insect cells, and birds, and of mammalian origin, e.g., human, primates, and rodents.

Prokaryotic host-vector systems include a wide variety of vectors for many different species. As used herein, E. coli and its vectors will be used generically to include equivalent vectors used in other prokaryotes. A representative vector for amplifying DNA is pBR322 or many of its derivatives. Vectors that can be used to express these chemokines or their fragments include, but are not limited to, such vectors as those containing the lac promoter (pUC-series); trp promoter (pBR322-trp); Ipp promoter (the pIN-series); lambda-pP or pR promoters (pOTS); or hybrid promoters such as ptac (pDR540). See Brosius, et al. (1988) "Expression Vectors Employing Lambda-, trp-, lac-, and Ipp-derived Promoters", in Rodriguez and Denhardt (eds.) Vectors: A Survey of Molecular Cloning Vectors and Their Uses, Buttersworth, Boston, Chapter 10, pp. 205-236.

Lower eukaryotes, e.g., yeasts and Dictyostelium, may be transformed with chemokine or receptor sequence containing nucleic acids. For purposes of this invention, the most common lower eukaryotic host is the baker's yeast, Saccharomyces cerevisiae. It will be used to generically represent lower eukaryotes although a number of other strains and species are also available. Yeast vectors typically consist of a replication origin (unless of the integrating type), a selection gene, a promoter, DNA encoding the desired protein or its fragments, and sequences for translation termination, polyadenylation, and transcription termination. Suitable expression vectors for yeast include such constitutive promoters as 3-

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phosphoglycerate kinase and various other glycolytic enzyme gene promoters or such inducible promoters as the alcohol dehydrogenase 2 promoter or metallothionine promoter. Suitable vectors include derivatives of the following types: self-replicating low copy number (such as the YRp-series), self-replicating high copy number (such as the YEp-series); integrating types (such as the YIp-series), or mini-chromosomes (such as the YCp-series).

Higher eukaryotic tissue culture cells are the preferred host cells for expression of the functionally active chemokine or receptor proteins. In principle, most any higher eukaryotic tissue culture cell line is workable, e.g., insect baculovirus expression systems, whether from an invertebrate or vertebrate source. However, mammalian cells are preferred, in that the processing, both cotranslationally and posttranslationally, will be typically most like natural. Transformation or transfection and propagation of such cells has become a routine procedure. Examples of useful cell lines include HeLa cells, Chinese hamster ovary (CHO) cell lines, baby rat kidney (BRK) cell lines, insect cell lines, bird cell lines, and monkey (COS) cell lines. Expression vectors for such cell lines usually include an origin of replication, a promoter, a translation initiation site, RNA splice sites (if genomic DNA is used), a polyadenylation site, and a transcription termination site. These vectors also usually contain a selection gene or amplification gene. Suitable expression vectors may be plasmids, viruses, or retroviruses carrying promoters derived, e.g., from such sources as from adenovirus, SV40, parvoviruses, vaccinia virus, or cytomegalovirus. Representative examples of suitable expression vectors include pCDNA1; pCD, see Okayama, et al. (1985) Mol. Cell Biol. 5:1136-1142; pMC1neo Poly-A, see Thomas, et al. (1987) Cell 51:503-512; and a baculovirus vector such as pAC 373 or pAC 610.

It will often be desired to express a chemokine or receptor polypeptide in a system which provides a specific or defined glycosylation pattern. In this case, the usual pattern will be that provided naturally by the expression system. However, the pattern will be modifiable by exposing the polypeptide, e.g., an unglycosylated form, to appropriate glycosylating proteins

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introduced into a heterologous expression system. For example, a chemokine or receptor gene may be co-transformed with one or more genes encoding mammalian or other glycosylating enzymes. Using this approach, certain mammalian glycosylation patterns will be achievable or approximated in prokaryote or other cells.

A chemokine, receptor, or a fragment thereof, may be engineered to be phosphatidyl inositol (PI) linked to a cell membrane, but can be removed from membranes by treatment with a phosphatidyl inositol cleaving enzyme, e.g., phosphatidyl inositol phospholipase-C. This releases the antigen in a biologically active form, and allows purification by standard procedures of protein chemistry. See, e.g., Low (1989) <u>Biochim. Biophys. Acta</u> 988:427-454; Tse, et al. (1985) <u>Science</u> 230:1003-1008; and Brunner, et al. (1991) <u>I. Cell Biol.</u> 114:1275-1283.

Now that these chemokines and receptors have been characterized, fragments or derivatives thereof can be prepared by conventional processes for synthesizing peptides. These include processes such as are described in Stewart and Young (1984) Solid Phase Peptide Synthesis, Pierce Chemical Co., Rockford, IL; Bodanszky and Bodanszky (1984) The Practice of Peptide Synthesis, Springer-Verlag, New York; and Bodanszky (1984) The Principles of Peptide Synthesis, Springer-Verlag, New York. For example, an azide process, an acid chloride process, an acid anhydride process, a mixed anhydride process, an active ester process (for example, pnitrophenyl ester, N-hydroxysuccinimide ester, or cyanomethyl ester), a carbodiimidazole process, an oxidative-reductive process, or a dicyclohexyl-carbodiimide (DCCD)/additive process can be used. Solid phase and solution phase syntheses are both applicable to the foregoing processes.

These chemokines, receptors, fragments, or derivatives are suitably prepared in accordance with the above processes as typically employed in peptide synthesis, generally either by a so-called stepwise process which comprises condensing an amino acid to the terminal amino acid, one by one in sequence, or by coupling peptide fragments to the terminal amino acid. Amino groups that are not

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being used in the coupling reaction are typically protected to prevent coupling at an incorrect location.

If a solid phase synthesis is adopted, the C-terminal amino acid is typically bound to an insoluble carrier or support through its carboxyl group. The insoluble carrier is not particularly limited as long as it has a binding capability to a reactive carboxyl group. Examples of such insoluble carriers include halomethyl resins, such as chloromethyl resin or bromomethyl resin, hydroxymethyl resins, phenol resins, tert-alkyloxycarbonyl-hydrazidated resins, and the like.

An amino group-protected amino acid is bound in sequence through condensation of its activated carboxyl group and the reactive amino group of the previously formed peptide or chain, to synthesize the peptide step by step. After synthesizing the complete sequence, the peptide is split off from the insoluble carrier to produce the peptide. This solid-phase approach is generally described, e.g., by Merrifield, et al. (1963) in <u>I. Am. Chem. Soc.</u> 85:2149-2156.

The prepared ligand and fragments thereof can be isolated and purified from the reaction mixture by means of peptide separation, e.g., by extraction, precipitation, electrophoresis, and various forms of chromatography, and the like. The various chemokines or receptors of this invention can be obtained in varying degrees of purity depending upon its desired use. Purification can be accomplished by use of the protein purification techniques disclosed herein or by the use of the antibodies herein described, e.g., in immunoabsorbant affinity chromatography. immunoabsorbant affinity chromatography is typically carried out, e.g., by first linking the antibodies to a solid support and then contacting the linked antibodies with solubilized lysates of appropriate source cells, lysates of other cells expressing the ligand or receptor, or lysates or supernatants of cells producing the desired proteins as a result of DNA techniques, see below.

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VIII. Uses

The present invention provides reagents which will find use in diagnostic applications as described elsewhere herein, e.g., in the general description for developmental abnormalities, or below in the description of kits for diagnosis.

This invention also provides reagents with significant therapeutic potential. These chemokines and receptors (naturally occurring or recombinant), fragments thereof, and binding compositions, e.g., antibodies thereto, along with compounds identified as having binding affinity to them, should be useful in the treatment of conditions associated with abnormal physiology or development, including inflammatory conditions, e.g., asthma. In particular, modulation of trafficking of leukocytes is one likely biological activity, but a wider tissue distribution might suggest broader biological activity, including, e.g., antiviral Abnormal proliferation, regeneration, degeneration, and atrophy may be modulated by appropriate therapeutic treatment using the compositions provided herein. For example, a disease or disorder associated with abnormal expression or abnormal signaling by a chemokine or ligand for a receptor should be a likely target for an agonist or antagonist of the ligand.

Various abnormal physiological or developmental conditions are known in cell types shown to possess the chemokine or receptor mRNAs by Northern blot analysis. See Berkow (ed.) The Merck Manual of Diagnosis and Therapy, Merck & Co., Rahway, N.J.; and Thorn, et al. Harrison's Principles of Internal Medicine, McGraw-Hill, N.Y. Developmental or functional abnormalities, e.g., of the immune system, cause significant medical abnormalities and conditions which may be susceptible to prevention or treatment using compositions provided herein.

Antibodies to the chemokines or receptors, including recombinant forms, can be purified and then used diagnostically or therapeutically, alone or in combination with other chemokines, cytokines, or antagonists thereof. These reagents can be combined for therapeutic use with additional active or inert ingredients, e.g., in conventional pharmaceutically acceptable carriers or diluents, e.g.,

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immunogenic adjuvants, along with physiologically innocuous stabilizers and excipients. These combinations can be sterile filtered and placed into dosage forms as by lyophilization in dosage vials or storage in stabilized aqueous preparations. This invention also contemplates use of antibodies or binding fragments thereof, including forms which are not complement binding. Moreover, modifications to the antibody molecules or antigen binding fragments thereof, may be adopted which affect the pharmacokinetics or pharmacodynamics of the therapeutic entity.

Drug screening using antibodies or receptor or fragments thereof can be performed to identify compounds having binding affinity to each chemokine or receptor, including isolation of associated components. Subsequent biological assays can then be utilized to determine if the compound has intrinsic stimulating activity and is therefore a blocker or antagonist in that it blocks the activity of the ligand. Likewise, a compound having intrinsic stimulating activity can activate the receptor and is thus an agonist in that it simulates the activity of a ligand. This invention further contemplates the therapeutic use of antibodies to these chemokines as antagonists, or to the receptors as antagonists or agonists. This approach should be particularly useful with other chemokine or receptor species variants.

The quantities of reagents necessary for effective therapy will depend upon many different factors, including means of administration, target site, physiological state of the patient, and other medicants administered. Thus, treatment dosages should be titrated to optimize safety and efficacy in various populations, including racial subgroups, age, gender, etc. Typically, dosages used in vitro may provide useful guidance in the amounts useful for in situ administration of these reagents. Animal testing of effective doses for treatment of particular disorders will provide further predictive indication of human dosage. Various considerations are described, e.g., in Gilman, et al. (eds.) (1990) Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th Ed., Pergamon Press; and Remington's Pharmaceutical Sciences, 17th ed. (1990), Mack Publishing Co., Easton, Penn.. Methods for administration are

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discussed therein and below, e.g., for oral, intravenous, intraperitoneal, or intramuscular administration, transdermal diffusion, and others. Pharmaceutically acceptable carriers typically include water, saline, buffers, and other compounds described, e.g., in the Merck Index, Merck & Co., Rahway, New Jersey. Dosage ranges would ordinarily be expected to be in amounts lower than 1 mM concentrations, typically less than about 10 µM concentrations, usually less than about 100 nM, preferably less than about 10 pM (picomolar), and most preferably less than about 1 fM (femtomolar), with an appropriate carrier. Slow release formulations, or a slow release apparatus often be utilized for continuous administration.

A chemokine, fragments thereof, or antibodies to it or its fragments, antagonists, and agonists, may be administered directly to the host to be treated or, depending on the size of the compounds, it may be desirable to conjugate them to carrier proteins such as ovalbumin or serum albumin prior to their administration. Therapeutic formulations may be administered in conventional dosage formulations. While it is possible for the active ingredient to be administered alone, it is often preferable to present it as a pharmaceutical formulation. Formulations typically comprise at least one active ingredient, as defined above, together with one or more acceptable carriers thereof. Each carrier should be both pharmaceutically and physiologically acceptable in the sense of being compatible with the other ingredients and not injurious to the Carriers may improve storage life, stability, etc. Formulations include those suitable for oral, rectal, nasal, or parenteral (including subcutaneous, intramuscular, intravenous and intradermal) administration. The formulations may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy. See, e.g., Gilman, et al. (eds.) (1990) Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th Ed., Pergamon Press; and Remington's Pharmaceutical Sciences, 17th ed. (1990), Mack Publishing Co., Easton, Penn.; Avis, et al. (eds.) (1993) Pharmaceutical Dosage Forms: Parenteral Medications Dekker, New York; Lieberman, et al. (eds.)

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(1990) <u>Pharmaceutical Dosage Forms: Tablets</u> Dekker, New York; and Lieberman, et al. (eds.) (1990) <u>Pharmaceutical Dosage Forms: Disperse Systems</u> Dekker, New York. The therapy of this invention may be combined with or used in association with other therapeutic agents. Similar considerations will often apply to receptor based reagents.

Both the naturally occurring and the recombinant forms of the chemokines or receptors of this invention are particularly useful in kits and assay methods which are capable of screening compounds for binding activity to the proteins. Several methods of automating assays have been developed in recent years so as to permit screening of tens of thousands of compounds in a short period. See, e.g., Fodor, et al. (1991) Science 251:767-773, which describes means for testing of binding affinity by a plurality of defined polymers synthesized on a solid substrate. The development of suitable assays can be greatly facilitated by the availability of large amounts of purified, soluble chemokine as provided by this invention.

For example, antagonists can normally be found once a ligand has been structurally defined. Testing of potential ligand analogs is now possible upon the development of highly automated assay methods using physiologically responsive cells. In particular, new agonists and antagonists will be discovered by using screening techniques described herein.

Viable cells could also be used to screen for the effects of drugs on respective chemokine or G-protein coupled receptor mediated functions, e.g., second messenger levels, i.e., Ca⁺⁺; inositol phosphate pool changes (see, e.g., Berridge (1993) Nature 361:315-325 or Billah and Anthes (1990) Biochem. J. 269:281-291); cellular morphology modification responses; phosphoinositide lipid turnover; an antiviral response. and others. Some detection methods allow for elimination of a separation step, e.g., a proximity sensitive detection system. Calcium sensitive dyes will be useful for detecting Ca⁺⁺ levels, with a fluorimeter or a fluorescence cell sorting apparatus.

Rational drug design may also be based upon structural studies of the molecular shapes of the chemokines, other effectors or analogs, or the receptors. Effectors may be other proteins which

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mediate other functions in response to ligand binding, or other proteins which normally interact with the receptor. One means for determining which sites interact with specific other proteins is a physical structure determination, e.g., x-ray crystallography or 2 dimensional NMR techniques. These will provide guidance as to which amino acid residues form molecular contact regions. For a detailed description of protein structural determination, see, e.g., Blundell and Johnson (1976) <u>Protein Crystallography</u>, Academic Press, New York.

Purified chemokine or receptor can be coated directly onto plates for use in the aforementioned drug screening techniques, and may be associated with detergents or lipids. However, non-neutralizing antibodies, e.g., to the chemokines or receptors can be used as capture antibodies to immobilize the respective protein on the solid phase.

Similar concepts also apply to the chemokine receptor embodiments of the invention.

IX. Kits

This invention also contemplates use of chemokine or receptor proteins, fragments thereof, peptides, binding compositions, and their fusion products in a variety of diagnostic kits and methods for detecting the presence of ligand, antibodies, or receptors. Typically the kit will have a compartment containing a defined chemokine or receptor peptide or gene segment or a reagent which recognizes one or the other, e.g., binding reagents.

A kit for determining the binding affinity of a test compound to a chemokine or receptor would typically comprise a test compound; a labeled compound, for example an antibody having known binding affinity for the protein; a source of chemokine or receptor (naturally occurring or recombinant); and a means for separating bound from free labeled compound, such as a solid phase for immobilizing the ligand or receptor. Once compounds are screened, those having suitable binding affinity to the ligand or receptor can be evaluated in suitable biological assays, as are well known in the art, to determine whether they act as agonists or

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antagonists to the receptor. The availability of recombinant chemokine or receptor polypeptides also provide well defined standards for calibrating such assays or as positive control samples.

A preferred kit for determining the concentration of, for example, a chemokine or receptor in a sample would typically comprise a labeled compound, e.g., antibody, having known binding affinity for the target, a source of ligand or receptor (naturally occurring or recombinant) and a means for separating the bound from free labeled compound, for example, a solid phase for immobilizing the chemokine or receptor. Compartments containing reagents, and instructions for use or disposal, will normally be provided.

Antibodies, including antigen binding fragments, specific for the chemokine or receptor, or fragments are useful in diagnostic applications to detect the presence of elevated levels of chemokine, receptor, and/or its fragments. Such diagnostic assays can employ lysates, live cells, fixed cells, immunofluorescence, cell cultures, body fluids, and further can involve the detection of antigens related to the ligand or receptor in serum, or the like. Diagnostic assays may be homogeneous (without a separation step between free reagent and antigen complex) or heterogeneous (with a separation step). Various commercial assays exist, such as radioimmunoassay (RIA), enzymelinked immunosorbent assay (ELISA), enzyme immunoassay (EIA), enzyme-multiplied immunoassay technique (EMIT), substratelabeled fluorescent immunoassay (SLFIA), and the like. example, unlabeled antibodies can be employed by using a second antibody which is labeled and which recognizes the primary antibody to a chemokine or receptor or to a particular fragment thereof. Similar assays have also been extensively discussed in the literature. See, e.g., Harlow and Lane (1988) Antibodies: A Laboratory Manual, CSH.

Anti-idiotypic antibodies may have similar uses to diagnose presence of antibodies against a chemokine or receptor, as such may be diagnostic of various abnormal states. For example, overproduction of a chemokine or receptor may result in production of various immunological reactions which may be diagnostic of

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abnormal physiological states, particularly in various inflammatory or asthma conditions.

Frequently, the reagents for diagnostic assays are supplied in kits, so as to optimize the sensitivity of the assay. For the subject invention, depending upon the nature of the assay, the protocol, and the label, either labeled or unlabeled antibody or labeled chemokine or receptor is provided. This is usually in conjunction with other additives, such as buffers, stabilizers, materials necessary for signal production such as substrates for enzymes, and the like. Preferably, the kit will also contain instructions for proper use and disposal of the contents after use. Typically the kit has compartments or containers for each useful reagent. Desirably, the reagents are provided as a dry lyophilized powder, where the reagents may be reconstituted in an aqueous medium providing appropriate concentrations of reagents for performing the assay.

The aforementioned constituents of the drug screening and the diagnostic assays may be used without modification or may be modified in a variety of ways. For example, labeling may be achieved by covalently or non-covalently joining a moiety which directly or indirectly provides a detectable signal. In any of these assays, the ligand, test compound, chemokine, receptor, or antibodies thereto can be labeled either directly or indirectly. Possibilities for direct labeling include label groups: radiolabels such as ¹²⁵I, enzymes (U.S. Pat. No. 3,645,090) such as peroxidase and alkaline phosphatase, and fluorescent labels (U.S. Pat. No. 3,940,475) capable of monitoring the change in fluorescence intensity, wavelength shift, or fluorescence polarization. Possibilities for indirect labeling include biotinylation of one constituent followed by binding to avidin coupled to one of the above label groups.

There are also numerous methods of separating bound from the free ligand, or alternatively bound from free test compound. The chemokine or receptor can be immobilized on various matrixes, perhaps with detergents or associated lipids, followed by washing. Suitable matrixes include plastic such as an ELISA plate, filters, and beads. Methods of immobilizing the chemokine or receptor to a matrix include, without limitation, direct adhesion to plastic, use of

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a capture antibody, chemical coupling, and biotin-avidin. The last step in this approach may involve the precipitation of antigen/antibody complex by any of several methods including those utilizing, e.g., an organic solvent such as polyethylene glycol or a salt such as ammonium sulfate. Other suitable separation techniques include, without limitation, the fluorescein antibody magnetizable particle method described in Rattle, et al. (1984) <u>Clin. Chem.</u> 30:1457-1461, and the double antibody magnetic particle separation as described in U.S. Pat. No. 4,659,678.

Methods for linking proteins or their fragments to the various labels have been extensively reported in the literature and do not require detailed discussion here. Many of the techniques involve the use of activated carboxyl groups either through the use of carbodiimide or active esters to form peptide bonds, the formation of thioethers by reaction of a mercapto group with an activated halogen such as chloroacetyl, or an activated olefin such as maleimide, for linkage, or the like. Fusion proteins will also find use in these applications.

Another diagnostic aspect of this invention involves use of oligonucleotide or polynucleotide sequences taken from the sequence of the chemokine or receptor. These sequences can be used as probes for detecting levels of the ligand message in samples from patients suspected of having an abnormal condition, e.g., an inflammatory, physiological, or developmental problem. preparation of both RNA and DNA nucleotide sequences, the labeling of the sequences, and the preferred size of the sequences has received ample description and discussion in the literature. Normally an oligonucleotide probe should have at least about 14 nucleotides, usually at least about 18 nucleotides, polynucleotide probes may be up to several kilobases. Various labels may be employed, most commonly radionuclides, particularly ³²P. However, other techniques may also be employed, such as using biotin modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescers, enzymes, or the like. Alternatively,

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antibodies may be employed which can recognize specific duplexes, including DNA duplexes, RNA duplexes, DNA-RNA hybrid duplexes, or DNA-protein duplexes. The antibodies in turn may be labeled and the assay carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected. The use of probes to the novel anti-sense RNA may be carried out in conventional techniques such as nucleic acid hybridization, plus and minus screening, recombinational probing, hybrid released translation (HRT), and hybrid arrested translation (HART). This also includes amplification techniques such as polymerase chain reaction (PCR).

Diagnostic kits which also test for the qualitative or quantitative presence of other markers are also contemplated. Diagnosis or prognosis may depend on the combination of multiple indications used as markers. Thus, kits may test for combinations of markers. See, e.g., Viallet, et al. (1989) <u>Progress in Growth Factor Res.</u> 1:89-97.

20 X. Receptor for Chemokine; Ligands for Receptors

Having isolated a ligand binding partner of a specific interaction, methods exist for isolating the counter-partner. See, Gearing, et al EMBO J. 8:3667-4676 or McMahan, et al. (1991) EMBO J. 10:2821-2832. For example, means to label a chemokine without interfering with the binding to its receptor can be determined. For example, an affinity label can be fused to either the amino- or carboxy-terminus of the ligand. An expression library can be screened for specific binding of chemokine, e.g., by cell sorting, or other screening to detect subpopulations which express such a binding component. See, e.g., Ho, et al. (1993) Proc. Nat'l Acad. Sci. 90:11267-11271. Alternatively, a panning method may be used. See, e.g., Seed and Aruffo (1987) Proc. Nat'l. Acad. Sci. 84:3365-3369.

With a receptor, means to identify the ligand exist. Methods for using the receptor, e.g., on the cell membrane, can be used to screen for ligand by, e.g., assaying for a common G-protein linked signal such as Ca++ flux. See Lerner (1994) Trends in Neurosciences

17:142-146. It is likely that the ligands for these receptors are chemokines.

Protein cross-linking techniques with label can be applied to a isolate binding partners of a chemokine. This would allow identification of protein which specifically interacts with a chemokine, e.g., in a ligand-receptor like manner.

The broad scope of this invention is best understood with reference to the following examples, which are not intended to limit the invention to specific embodiments.

EXAMPLES

I. General Methods

Some of the standard methods are described or referenced, 15 e.g., in Maniatis, et al. (1982) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor Press; Sambrook, et al. (1989) Molecular Cloning: A Laboratory Manual, (2d ed.), vols 1-3, CSH Press, NY; Ausubel, et al., Biology, Greene Publishing Associates, Brooklyn, NY; or Ausubel, et al. (1987 and 20 Supplements) Current Protocols in Molecular Biology, Greene/Wiley, New York; Innis, et al. (eds.)(1990) PCR Protocols: A Guide to Methods and Applications Academic Press, N.Y. Methods for protein purification include such methods as ammonium sulfate chromatography, precipitation, column electrophoresis, centrifugation, crystallization, and others. See, e.g., Ausubel, et al. 25 (1987 and periodic supplements); Deutscher (1990) "Guide to Protein Purification" in Methods in Enzymology, vol. 182, and other volumes in this series; and manufacturer's literature on use of protein purification products, e.g., Pharmacia, Piscataway, N.J., or 30 Bio-Rad, Richmond, CA. Combination with recombinant techniques allow fusion to appropriate segments, e.g., to a FLAG sequence or an equivalent which can be fused via a proteaseremovable sequence. See, e.g., Hochuli (1989) Chemische Industrie 12:69-70; Hochuli (1990) "Purification of Recombinant Proteins with 35 Metal Chelate Absorbent" in Setlow (ed.) Genetic Engineering, Principle and Methods 12:87-98, Plenum Press, N.Y.; and Crowe, et

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al. (1992) <u>OIAexpress: The High Level Expression & Protein Purification System</u> QUIAGEN, Inc., Chatsworth, CA.

FACS analyses are described in Melamed, et al. (1990) Flow Cytometry and Sorting Wiley-Liss, Inc., New York, NY; Shapiro (1988) Practical Flow Cytometry Liss, New York, NY; and Robinson, et al. (1993) Handbook of Flow Cytometry Methods Wiley-Liss, New York, NY.

II. Isolation and characterization of chemokine cDNAs

A. Primate IBICK

The IBICK was isolated from a cDNA library made from a human astrocytoma cell line. See, e.g., Rani, et al. (1996) <u>I. Biol. Chem.</u> 271:22878-22884. There is reported a gene which is not inducible by IFN- α , but inducible by IFN- γ and/or IFN- β . Applicants have identified this gene as a chemokine, and designated it Interferon Beta Induced ChemoKine (IBICK), which is described in SEQ ID NO: 2. Individual cDNA clones are sequenced using standard methods, e.g., the Taq DyeDeoxy Terminator Cycle Sequencing kit (Applied Biosystems, Foster City, CA), and the sequence is further characterized.

The predicted signal sequence corresponds to amino acids met1 to about gly21, so the mature form should begin with phe22 and run about 74 amino acids. Additional processing may occur in a physiological system.

Computer analysis and alignments for related genes indicates the closest match is to the two IFN- γ regulated chemokines MIG and IP10, but other related molecules are chemoknes. See, e.g., Faubert (1993) <u>Biochem. Biophys. Res. Commun.</u> 192:223-230; and Luster, et al. (1985) <u>Nature 315:672-676</u>. This similarity in sequence may well correlate with similarity in regulation, which suggests related functions. The rarity of related sequences in the existing sequence databases suggests low message levels, tight negative regulation, and/or a distribution pattern in cell types not yet analysed. The IFN- γ regulatable nature of this chemokine suggests a role as an antiviral or antitumor agent. Its non-ELR chemokine structure suggests

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angiostatic, in contrast to angiogenic, activity, which may be important in tumor therapy.

Other primate counterparts should be isolatable using the entire coding portion of this human clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

B. Primate ILINCK

The ILINCK (SEQ ID NO: 3 and 4) was isolated from a cDNA library made from a human liver cell library. Total RNA can be isolated, e.g., using the guanidine thiocyanate/CsCl gradient procedure as described by Chirgwin, et al. (1978) <u>Biochem.</u> 18:5294-5299. Poly(A)+ RNA is isolated using, e.g., the OLIGOTEX mRNA isolation kit (QIAGEN). Such RNA from these cells is used to synthesize first strand cDNA, e.g., by using NotI/Oligo-dT primer (Gibco-BRL, Gaithersburg, MD). Double-stranded cDNA is synthesized, ligated with BstXI adaptors, digested with NotI, size fractionated for > 0.5 kilobase pairs (kb) and ligated into the NotI/BstXI sites of pJFE-14, a derivative of the pCDSRa vector. See Takebe, et al. (1985) <u>Mol. Cell Biol.</u> 8:466-472. Electro-competent E. coli DH10a cells (Gibco-BRL) are used for transformation.

The gene apparently produces at least two different sized transcripts, 0.6 kB and a 1.5 kB, which are differently regulated. The larger transcript is inducible by IL-10, which is unusual for a chemokine, so it has been designated InterLeukin 10 INduced ChemoKine (ILINCK), which is described in SEQ ID NO: 4. Individual cDNA clones were sequenced using standard methods, e.g., the Taq DyeDeoxy Terminator Cycle Sequencing kit (Applied Biosystems, Foster City, CA), and the sequence was further characterized.

The predicted signal sequence corresponds to amino acids metl to about ser23, so the mature form should begin with gln24 and run about 73 amino acids. Additional processing may occur in a physiological system.

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Computer analysis and alignments for related genes indicates the closest match is to other chemokines.

Other primate counterparts should be isolatable using the entire coding portion of this human clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

The ILINCK mRNA is induced in monocytes by IL-10, a most notable feature. This observation strongly suggests that ILINCK has anti-inflammatory properties. It is postulated that ILINCK will be a potential therapeutic in autoimmune or other inflammatory disorders. See, e.g., Samter, et al. (eds) <u>Immunological Diseases</u> vols. 1 and 2, Little, Brown and Co.

C. Rodent CXC-143

The CXC-143 (SEQ ID NO: 5) was isolated from a cDNA library made from a mouse placenta cDNA library. The partial sequence provided lacks an identifiable initiation codon and termination codon. This chemokine has been designated CXC-143, and is described in SEQ ID NO: 6, 8 and 10. Individual cDNA clones were sequenced using standard methods, e.g., the Taq DyeDeoxy Terminator Cycle Sequencing kit (Applied Biosystems, Foster City, CA), and the sequence was further characterized, but the sequence remains incomplete. Clearly the chemokine is a non-ELR class CXC chemokine.

Computer analysis and alignments for related genes indicates the closest match is to other chemokine, the MIG, IP10, and the IBICK, all of which are IFN- γ inducible. This sequence similarity suggests a similar transcriptional regulation, and similar uses to the IBICK described above.

Other primate counterparts should be isolatable using the entire coding portion of this human clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

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D. Rodent MCP243

The MCP243 (SEQ ID NO: 11) was isolated from a cDNA library made from a mouse cDNA library. The partial sequence provided lacks an identifiable initiation codon and various upstream chemokine motifs. This chemokine has been designated MCP23, and is described in SEQ ID NO: 12 and 14. Individual cDNA clones are sequenced using standard methods, e.g., the Taq DyeDeoxy Terminator Cycle Sequencing kit (Applied Biosystems, Foster City, CA), and the sequence further characterized, but the sequence remains incomplete.

Computer analysis and alignments for related genes indicates the closest match is to various monocyte chemoattractant proteins. Clearly the encoded protein is a chemokine, and will have many similar biological activities related to the other members of the MCPs.

Other rodent counterparts should be isolatable using the entire coding portion of this mouse clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

III. Isolation and characterization of GPCR cDNAs

A. Primate R277

The primate R277 clone was derived from human fetal tissue cDNA library. The nucleotide and amino acid sequences are provided in SEQ ID NO: 15, 16, 17 and 18.

Computer analysis suggests that the closest related genes are various G-protein coupled receptors. These include the chemokine receptors, and protease, e.g., thrombin, receptors. Structural motifs suggest that the receptor may contain motifs characteristic of the chemokine receptor family, and of the protease receptor family. The transmembrane segments, based upon hydrophobicity plots and comparisons with other similar GPCRs, should be about as follows for SEQ ID NO: 16: TM3 to val17; TM4 from arg36 to leu57; TM5 from asn89 to arg111; TM6 from leu134 to leu161; and TM7 from met178 to val200.

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See, e.g., Loetscher, et al. (1996) <u>I. Expt'l Med.</u> 184:963-969. A DRY motif is found, e.g., near residue 18. The amino terminal segment is probably an extracellular segment (E1), and the others would be E2 between TM2 and TM3; E3 between TM4 and TM5; and E4 between TM6 and TM7. The intracellular segments should then run I1 between TM1 and TM2; I2 between TM3 and TM4, I3 between TM5 and TM6, and I4 the carboxy terminus from the end of TM7. Additional processing may occur in a physiological system. Conserved residues among the GPCRs would include, e.g., arg33, cys67, and gly217, among others in the transmembrane segments. A computer analysis of GPCR sequences will indicate residues characteristic of the family members. Core transmembrane segments for the R277 receptor sequence are predicted, using SEQ ID NO: 18 numbering, about: TM1 leu16 to leu41; TM2 ile51 to ile71; TM3 leu93 to leu125; TM4 leu132 to leu150; TM5 leu183 to val206; TM6 ile224 to ala256; and TM7 ile274 to val293.

Other primate counterparts should be isolatable using the entire coding portion of this human clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

B. Rodent HST01.1

The rodent HST01.1 clone was derived from a cDNA library made from mouse TcR ab+ CD4- CD8- T cells. See Zlotnik, et al. (1992) <u>I. Immunol</u>. 149:1211-1215. Individual cDNA clones are sequenced using standard methods, and the sequence identified and further characterized. The partial nucleotide sequence is provided in SEQ ID NO: 19, encoding a polypeptide fragment of about 74 amino acids (SEQ ID NO: 20). Complete rodent HSTO1.1 nucleotide and amino acid sequences are provided in SEQ ID NO: 21 and 22.

Computer analysis suggests that the closest related genes are various G-protein coupled receptors. Structural motifs suggest that the receptor may contain motifs characteristic of the chemokine receptor family, and of the protease receptor family. In SEQ ID NO: 22, the transmembrane segments, based upon hydrophobicity plots and comparisons with other similar GPCRs, should be about as

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follows: TM1 from leu58 to leu78; TM2 from phe90 to val110; TM3 from ala126 to phe146; TM4 from leu169 to leu189; TM5 from phe223 to val243; TM6 from val256 to asp277; and TM7 from val301 to fly321. A DRY box motif runs from about asp147 to ala155. Chemokine receptors are generally considered useful targets for novel drug discovery, where the therapeutics would agonise or antagonise the binding of natural ligand(s) of the receptor. These receptor-ligand interactions may result in inflammation, cell recruitment, an/or cell activation processes. Some of these receptors are the portal of entry of infectious agents, e.g., viruses. Therefore, therapeutics directed against the chomokine receptor may find application in these diseases. In addition, the receptors may be important in determining fundamental structure or physiological responses.

Other rodent counterparts should be isolatable using the entire coding portion of this mouse clone as a hybridization probe. A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

Ligand-receptor analysis has indicated that this receptor, when transfected into a cell, makes that cell responsive to the presence of chemokines IP-10 (IFN-γ-Inducible Protein-10), (monokine induced by IFN-g), and 6Ckine. See, e.g., Luster, et al. (1985) Nature 315:672-676; Ohmori and Hamilton (1990) Biochem. Biophys. Res. Commun. 168:1261-1267; Vanguri and Farber (1990) J. Biol. Chem. 265:15049-15057; Farber (1990) Proc. Nat'l Acad. Sci. USA 87:5238-5242; Farber (1993) Biochem. Biophys. Res. Commun. 192:223-230; and GenBank accession numbers AF006637; U88320, and U88322. However, the 6Ckine seems to bind differently from the MIG and IP-10, as it is incapable of desensitizing the response of the receptor to the other chemokines. MIG can desensitize the response to 6Ckine, but IP-10 does not. These results imply that 6Ckine may have angiostatic and antitumor activities similar to those of MIG See, e.g., Sgadari, et al. (1997) Blood 89:2635-2643; Arenberg, et al. (1996) J. Exp. Med. 184:981-992; Loetscher, et al. (1996)

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J. Exp. Med. 184:963-969; Sgadari, et al. (1996) Proc. Natl Acad. Sci. USA 93:13791-13796; Angiolillo, et al. (1996) Ann. NY Acad. Sci. 795:158-167; Sarris, et al. (1996) Leukemia 10:757-765; Angiolillo, et al. (1995) J. Exp. Med. 182:155-162; Strieter, et al. (1995) J. Leukoc. Biol. 57:752-762; Strieter, et al. (1995) Biochem. Biophys. Res. Commun. 210:51-57; Clark-Lewis, et al. (1994) J. Biol. Chem. 269:16075-16081.

C. Rodent 941D12

The rodent 941D12 clone was derived from a cDNA library made from mouse Th3 polarized cells. Production of 3W Th1 or Th2 cells is described in Openshaw, et al. (1995) J. Exp. Med. 182:1357-1367. Briefly, Th1 or Th2 populations were derived from CD4+ T cells stimulated with antigen and antigen presenting cells in the presence of IL-12 or IL-4. Cells were stimulated once each week for 3 weeks, then harvested and restimulated, e.g., with PMA and ionmycin for 4 h. See Murphy, et al. (1996) J. Exp. Med. 183:901-913. A subtraction step was introduced to remove sequences found in mouse L cells. See, e.g., Hara, et al. (1994) Blood 84:189-199.

Individual cDNA clones are sequenced using standard methods, and the sequence identified and further characterized. The partial nucleotide sequence is provided in SEQ ID NO: 23, encoding a polypeptide fragment of about 193 amino acids (SEQ ID NO: 24). Complete rodent 94ID12 nucleotide and amino acid sequences are provided in SEQ ID NO: 25 and 26.

Computer analysis suggests that the closest related genes are various G-protein coupled receptors. Structural motifs suggest that the receptor may contain motifs characteristic of the chemokine receptor family, and of the protease receptor family. The transmembrane segments, based upon hydrophobicity plots and comparisons with other similar GPCRs, should be about as follows on SEQ ID NO: 24: TM1 from val62 to phe88; TM2 from val98 to ala120; TM3 from val145 to leu157. For SEQ ID NO: 26, the predicted core transmembrane segments are about TM1 leu48 to ile64; TM2 val77 to leu93; TM3 val124 to val140; TM4 ile155 to val171; TM5 leu201 to ile217; TM6 ser238 to thr254; and TM7 ile279 to leu295.

Other rodent counterparts should be isolatable using the entire coding portion of this mouse clone as a hybridization probe.

A Southern blot may indicate the extent of homology across species, and either a cDNA library or mRNA can be screened to identify an appropriate cell source. The physiological state of many different cell types may also be evaluated for increased expression of the gene.

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IV. Preparation of antibodies

Many standard methods are available for preparation of antibodies. For example, synthetic peptides may be prepared to be used as antigen, administered to an appropriate animal, and either polyclonal or monoclonal antibodies prepared. Short peptides, e.g., less than about 10 amino acids may be expressed as repeated sequences, while longer peptides may be used alone or conjugated to a carrier. For example, with the GPCRs, animals are immunized with peptides or complete proteins from SEQ ID NO: 12, 14, 16, or 18. Highest specificity will result when the polypeptides are selected from portions which are most unique, e.g., not from conserved sequence regions. The animals may be used to collect antiserum, or may be used to generate monoclonal antibodies.

Antiserum is evaluated for use, e.g., in an ELISA, and will be evaluated for utility in immunoprecipitation, e.g., typically native, or Western blot, e.g., denatured antigen, analysis. Monoclonal antibodies will also be evaluated for those same uses.

The antibodies provided will be useful as immunoaffinity reagents, as detection reagents, for immunohistochemistry, and as potential therapeutic reagents, either as agonist or antagonist reagents.

V. Assays for chemotactic activity of chemokines

Chemokine proteins are produced, e.g., in COS cells transfected with a plasmid carrying the chemokine cDNA by electroporation. See, Hara, et al. (1992) EMBO J. 10:1875-1884. Physical analytical methods may be applied, e.g., CD analysis, to compare tertiary structure to other chemokines to evaluate whether the protein has likely folded into an active conformation. After transfection, a culture supernatant is collected and subjected to bioassays. A mock control, e.g., a plasmid carrying the luciferase

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cDNA, is used. See, de Wet, et al. (1987) Mol. Cell. Biol. 7:725-757. A positive control, e.g., recombinant murine MIP-1a from R&D Systems (Minneapolis, MN), is typically used. Likewise, antibodies may be used to block the biological activities, e.g., as a control.

Lymphocyte migration assays are performed as previously described, e.g., in Bacon, et al. (1988) Br. J. Pharmacol. 95:966-974. Murine Th2 T cell clones, CDC-25 (see Tony, et al. (1985) J. Exp. Med. 161:223-241) and HDK-1 (see Cherwinski, et al. (1987) J. Exp. Med. 166:1229-1244), made available from R. Coffman and A. O'Garra (DNAX, Palo Alto, CA), respectively, are used as controls.

Ca2+ flux upon chemokine stimulation is measured, e.g., according to the published procedure described in Bacon, et al. (1995) <u>J. Immunol.</u> 154:3654-3666.

Maximal numbers of migrating cells in response to the IBICK are measured. See Schall (1993) J. Exp. Med. 177:1821-1826. A doseresponse curve is determined, preferably giving a characteristic bell shaped dose-response curve.

After stimulation with various chemokines, lymphocytes often exhibit a measurable intracellular Ca2+ flux. MIP-1a, e.g., is capable of inducing immediate transients of calcium mobilization. Typically, the levels of chemokine used in these assays will be comparable to those used for the chemotaxis assays (1/1000 dilution of conditioned supernatants).

Retroviral infection assays have also been described, and recent description of certain chemokine receptors in retroviral infection processes may indicate that similar roles may apply these See, e.g., Balter (1996) Science 272:1740 (describing evidence for chemokine receptors as coreceptors for HIV); and Deng, et al. (1996) Nature 381:661-666.

For receptors, biological activity may be measured in response to an appropriate ligand. The receptors are transfected into an assortment of cell types, each of which is likely to possess the intracellular signaling components compatible with the expressed receptor. Various ligand sources are tested to find a source of ligand 35 which results in a G-protein coupled response. Alternatively, the cells are tested for Ca++ flux in response to such ligands. Flux may

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be conveniently measured by electrophysiological means, or by Ca++ sensitive dyes.

VI. Analysis of individual variation

From the distribution data, an abundant easily accessible cell type is selected for sampling from individuals. Using PCR techniques, a large population of individuals are analysed for this gene. cDNA or other PCR methods are used to sequence the corresponding gene in the different individuals, and their sequences are compared. This indicates both the extent of divergence among racial or other populations, as well as determining which residues are likely to be modifiable without dramatic effects on function.

VII. Biological activities, direct and indirect

A robust and sensitive assay is selected as described above, e.g., on immune cells, neuronal cells, or stem cells. Chemokine is added to the assay in increasing doses to see if a dose response is detected. For example, in a proliferation assay, cells are plated out in plates. Appropriate culture medium is provided, and chemokine is added to the cells in varying amounts. Growth is monitored over a period of time which will detect either a direct effect on the cells, or an indirect effect of the chemokine.

Alternatively, an activation assay or attraction assay is used. An appropriate cell type is selected, e.g, hematopoietic cells, myeloid (macrophages, neutrophils, polymorphonuclear cells, etc.) or lymphoid (T cell, B cell, or NK cells), neural cells (neurons, neuroglia, oligodendrocytes, astrocytes, etc.), or stem cells, e.g., progenitor cells which differentiate to other cell types, e.g., gut crypt cells and undifferentiated cell types.

Other assays will be those which have been demonstrated with other chemokines. See, e.g., Schall and Bacon (1994) <u>Current Opinion in Immunology</u> 6:865-873; and Bacon and Schall (1996) <u>Int. Arch. Allergy & Immunol.</u> 109:97-109.

35 VIII. Structure activity relationship

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Information on the criticality of particular residues is determined using standard procedures and analysis. Standard mutagenesis analysis is performed, e.g., by generating many different variants at determined positions, e.g., at the structural positions identified above, and evaluating biological activities of the variants. This may be performed to the extent of determining positions which modify activity, or to focus on specific positions to determine the residues which can be substituted to either retain, block, or modulate biological activity.

Alternatively, analysis of natural variants can indicate what positions tolerate natural mutations. This may result from populational analysis of variation among individuals, or across strains or species. Samples from selected individuals are analyzed, e.g., by PCR analysis and sequencing. This allows evaluation of population polymorphisms.

IX. Chromosomal localization

The cDNA is labeled, e.g., nick-translated with biotin-14 dATP and hybridized in situ at a final concentration of 5 ng/ μ l to metaphases from two normal males. Fluorescence in situ hybridization (FISH) method may be modified from that described by Callen, et al. (1990). <u>Ann. Genet.</u> 33:219-221, in that chromosomes are stained before analysis with both prodidium iodide (as counter stain) and DAPI (for chromosome identification). Images of metaphase preparations are captured by a CCD camera and computer enhanced. Identification of the approapriate labeled chromosomes is determined.

X. Expression analysis of chemokine/receptor genes

RNA blot and hybridization are performed according to the standard methods in Maniatis, et al. (1982) Molecular Cloning: A Laboratory Manual Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY. An appropriate fragment or the whole coding sequence of a cDNA fragment is selected for use as a probe. To verify the amount of RNA loaded in each lane, the substrate membrane is

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reprobed with a control cDNA, e.g., glyceraldehyde 3-phosphate dehydrogenase (G3PDH) cDNA (Clontech, Palo Alto CA).

Analysis of mRNA from the appropriate cell source using the probe will determine the natural size of message. It will also indicate whether different sized messages exist. The messages will be subject to analysis after isolation, e.g., by PCR or hybridization techniques.

Northern blot analysis may be performed on many different mRNA sources, e.g., different tissues, different species, or cells exhibiting defined physiological responses, e.g., activation conditions or developmental conditions. However, in certain cases, cDNA libraries may be used to evaluate sources which are difficult to prepare. A "reverse Northern" uses cDNA inserts removed from vector, but multiplicity of bands may reflect either different sized messages, or may be artifact due to incomplete reverse transcription in the preparation of the cDNA library. In such instances, verification may be appropriate by standard Northern analysis.

Similarly, Southern blots may be used to evaluate species distribution of a gene. The stringency of washes of the blot will also provide information as to the extent of homology of various species counterparts.

Tissue distribution, and cell distribution, may be evaluated by immunohistochemistry using antibodies. Alternatively, in situ nucleic acid hybridization may also be used in such analysis. Certain distribution data may be ascertained by the frequency and tissue types where messages have been found and collected in sequence databases, e.g., GenBank or proprietary collections.

A. IBICK

The IBICK was isolated from a human astrocyte cell. There is little distribution data generated at this time.

B. ILINCK

The IBICK was isolated from a human liver library. It is expressed in NK cells, gd T cells, and activated and resting monocytes. Libraries from human T cell lines, e.g., Mot81, HY106, and Mut72 show expression. Northern blots of adult spleen, thymus, prostate, testis, yterus, small intestine, colon, and peripheral

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blood leukocytes give no detectable signal. These data suggest that the expression is in a limited subset of specialized cells. Neither activated splenocytes not activated PBLs give detectable signals.

C. CXC-143

The CXC-143 gene was identified from a cDNA library made from mouse placenta. The sequence has appeared in cDNA libraries from varous C57BL/6J mouse embryo or placenta. Notably, early embryo libraries, e.g., from 8.5 days post conception, exhibited signal. A homologous, human gene is found in placenta, fetal heart, breast, and fetal liver/spleen. This distibution suggests a role of the molecule in early stages of development. The molecule, or its antagonist, should be useful in various early developmental conditions.

D. R277 GPCR

The R277 gene was identified from a cDNA library made from 20 week human fetal liver/spleen tissues. Southern analysis in cDNA Libraries showed that the gene is highly expressed CD34 derived dendritic cells; and is also expressed in T & B cell libraries (varying degrees); activated monocytes; and NKL clone. On a Clontech Multiple Human Tissue blot, a transcript of about 2.4 kb was detected in spleen and PBL.

E. HST01.1 GPCR

The HST01.1 gene was isolated from an ab TCR+ CD4+ CD8+ cell library. Distribution analysis showed a strong positive signal by Northern analysis in spleen and lung. Southern analysis showed strong positive signals in Th1 clones, CD4+NK1.1+ cells, abTcR double negative cells, D1.1 resting Th1 T cells, and mesentary lymph nodes. Positive signals were detected in macrophages, Th2 cells, and CD3+abTcR double negative cells, D1.1 ConA stimulated Th1 T cells, resting J774 cells, and LPS and IL-10 stimulated J774 cells. Weaker signals were detected in thymus, activated pro-T cells, and LPS and anti-IL-10 treated J774 cells. Very weak signals were detected in large B cell line from spleen, from dendritic cells from a resting spleen, and colon. No signal was detected in resting pro-T cells, CD44-CD25+ pre-T cells, CD35+ resting Th2 T cells, ConA stimulated CD35+ Th2 T cells, mature B cell leukemia, CH12 B cell line, B cells

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from LPS treated spleen, resting dendritic cells from bone marrow, RAW264.7 moncyte cell line, and total Peyer's patch.

F. 941D12 GPCR

The 941D12 GPCR was isolated from a mouse 3 week polarized Th2 cell cDNA library, subtracted with cDNA sequences from a mouse L cell. See, e.g., Openshaw, et al. (1995) <u>J. Exp. Med.</u> 182:1357-1367; and Murphy, et al. (1996) <u>J. Exp. Med.</u> 183:901-913.

Southern Analysis: DNA (5 mg) from a primary amplified cDNA library was digested with appropriate restriction enzymes to release the inserts, run on a 1% agarose gel and transferred to a nylon membrane (Schleicher and Schuell, Keene, NH).

Samples for mouse mRNA isolation include, e.g.: resting mouse fibroblastic L cell line (C200); Braf:ER (Braf fusion to estrogen receptor) transfected cells, control (C201); T cells, TH1 polarized (Mel14 bright, CD4+ cells from spleen, polarized for 7 days with IFNg and anti IL-4; T200); T cells, TH2 polarized (Mel14 bright, CD4+ cells from spleen, polarized for 7 days with IL-4 and anti-IFN-y; T201); T cells, highly TH1 polarized (see Openshaw, et al. (1995) L Exp. Med. 182:1357-1367; activated with anti-CD3 for 2, 6, 16 h pooled; T202); T cells, highly TH2 polarized (see Openshaw, et al. (1995) J. Exp. Med. 182:1357-1367; activated with anti-CD3 for 2, 6, 16 h pooled; T203); CD44- CD25+ pre T cells, sorted from thymus (T204); TH1 T cell clone D1.1, resting for 3 weeks after last stimulation with antigen (T205); TH1 T cell clone D1.1, 10 mg/ml ConA stimulated 15 h (T206); TH2 T cell clone CDC35, resting for 3 weeks after last stimulation with antigen (T207); TH2 T cell clone CDC35, 10 mg/ml ConA stimulated 15 h (T208); Mel 14+ naive T cells from spleen, resting (T209); Mel14+ T cells, polarized to Th1 with IΦN-γ/IL-12/anti-IL-4 for 6, 12, 24 h pooled (T210); Mel 14+ T cells, polarized to Th2 with IL-4/anti-IΦN-γ for 6, 13, 24 h pooled (T211); unstimulated mature B cell leukemia cell line A20 (B200); unstimulated B cell line CH12 (B201); unstimulated large B cells from spleen (B202); B cells from total spleen, LPS activated (B203); metrizamide enriched dendritic cells from spleen, resting (D200); dendritic cells from bone marrow, resting (D201); monocyte cell line RAW 264.7 activated with LPS 4 h (M200); bone-marrow macrophages derived with GM

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and M-CSF (M201); macrophage cell line J774, resting (M202); macrophage cell line J774 + LPS + anti-IL-10 at 0.5, 1, 3, 6, 12 h pooled (M203); macrophage cell line J774 + LPS + IL-10 at 0.5, 1, 3, 5, 12 h pooled(M204); aerosol challenged mouse lung tissue, Th2 primers, aerosol OVA challenge 7, 14, 23 h pooled (see Garlisi, et al. (1995) Clinical Immunology and Immunopathology 75:75-83; X206); Nippostrongulus-infected lung tissue (see Coffman, et al. (1989) Science 245:308-310; X200); total adult lung, normal (O200); total lung, rag-1 (see Schwarz, et al. (1993) Immunodeficiency 4:249-252; O205); IL-10 K.O. spleen (see Kuhn, et al. (1991) Cell 75:263-274; X201); total adult spleen, normal (O201); total spleen, rag-1 (O207); IL-10 K.O. Peyer's patches (O202); total Peyer's patches, normal (O210); IL-10 K.O. mesenteric lymph nodes (X203); total mesenteric lymph nodes, normal (O211); IL-10 K.O. colon (X203); total colon, normal (O212); NOD mouse pancreas (see Makino, et al. (1980) <u>Jikken Dobutsu</u> 29:1-13; X205); total thymus, rag-1 (O208); total kidney, rag-1 (O209); total heart, rag-1 (O202); total brain, rag-1 (O203); total testes, rag-1 (O204); total liver, rag-1 (O206); rat normal joint tissue (O300); and rat arthritic joint tissue (X300).

High signals were detected in IL-10 K.O. Peyer's patches (O202); total Peyer's patches, normal (O210); TH2 T cell clone CDC35, resting for 3 weeks after last stimulation with antigen (T207); T cells, highly TH2 polarized (see Openshaw, et al. (1995) J. Exp. Med. 182:1357-1367; activated with anti-CD3 for 2, 6, 16 h pooled; T203); total kidney, rag-1 (O209); and total heart, rag-1 (O202). Significant signals were detected in bone-marrow macrophages derived with GM and M-CSF (M201); T cells, TH2 polarized (Mel14 bright, CD4+ cells from spleen, polarized for 7 days with IL-4 and anti-IΦN-γ; T201); dendritic cells from bone marrow, resting (D201); total brain, rag-1 (O203); total liver, rag-1 (O206); total colon, normal (O212); and total thymus, rag-1 (O208). Weak signals were detected in TH2 T cell clone CDC35, 10 mg/ml ConA stimulated 15 h (T208); macrophage cell line J774, resting (M202); IL-10 K.O. colon (X203); Mel 14+ T cells, polarized to Th2 with IL-4/anti-IFN-γ for 6, 13, 24 h pooled (T211); macrophage cell line J774 + LPS + anti-IL-10 at 0.5, 1, 3, 6, 12 h pooled (M203); TH1 T cell clone D1.1, 10 mg/ml ConA stimulated 15 h (T206); total lung,

rag-1 (see Schwarz, et al. (1993) <u>Immunodeficiency</u> 4:249-252; O205); total spleen, rag-1 (O207); IL-10 K.O. mesenteric lymph nodes (X203); total testes, rag-1 (O204); T cells, TH1 polarized (Mel14 bright, CD4+cells from spleen, polarized for 7 days with IΦN–γ and anti IL-4; T200); TH1 T cell clone D1.1, resting for 3 weeks after last stimulation with antigen (T205); monocyte cell line RAW 264.7 activated with LPS 4 h (M200); and Nippostrongulus-infected lung tissue (see Coffman, et al. (1989) <u>Science</u> 245:308-310; X200). Other samples in the list gave no detectable signal.

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XI. Screening for receptor/ligand

Labeled reagent is useful for screening of an expression library made from a cell line which expresses a chemokine or receptor, as appropriate. Standard staining techniques are used to detect or sort intracellular or surface expressed ligand, or surface expressing transformed cells are screened by panning. Screening of intracellular expression is performed by various staining or immunofluorescence procedures. See also, e.g., McMahan, et al. (1991) <u>EMBO I.</u> 10:2821-2832.

For example, on day 0, precoat 2-chamber permanox slides with 1 ml per chamber of fibronectin, 10 ng/ml in PBS, for 30 min at room temperature. Rinse once with PBS. Then plate COS cells at 2-3 x 10^5 cells per chamber in 1.5 ml of growth media. Incubate overnight at 37° C.

On day 1 for each sample, prepare 0.5 ml of a solution of 66 mg/ml DEAE-dextran, 66 mM chloroquine, and 4 mg DNA in serum free DME. For each set, a positive control is prepared, e.g., of huIL-10-FLAG cDNA at 1 and 1/200 dilution, and a negative mock. Rinse cells with serum free DME. Add the DNA solution and incubate 5 hr at 37° C. Remove the medium and add 0.5 ml 10% DMSO in DME for 2.5 min. Remove and wash once with DME. Add 1.5 ml growth medium and incubate overnight.

On day 2, change the medium. On days 3 or 4, the cells are fixed and stained. Rinse the cells twice with Hank's Buffered Saline Solution (HBSS) and fix in 4% paraformaldehyde (PFA)/glucose for 5 min. Wash 3X with HBSS. The slides may be stored at -80° C after

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all liquid is removed. For each chamber, 0.5 ml incubations are performed as follows. Add HBSS/saponin(0.1%) with 32 ml/ml of 1M NaN3 for 20 min. Cells are then washed with HBSS/saponin 1X. Add antibody complex to cells and incubate for 30 min. Wash cells twice with HBSS/saponin. Add second antibody, e.g., Vector antimouse antibody, at 1/200 dilution, and incubate for 30 min. Prepare ELISA solution, e.g., Vector Elite ABC horseradish peroxidase solution, and preincubate for 30 min. Use, e.g., 1 drop of solution A (avidin) and 1 drop solution B (biotin) per 2.5 ml HBSS/saponin. Wash cells twice with HBSS/saponin. Add ABC HRP solution and incubate for 30 min. Wash cells twice with HBSS, second wash for 2 min, which closes cells. Then add Vector diaminobenzoic acid (DAB) for 5 to 10 min. Use 2 drops of buffer plus 4 drops DAB plus 2 drops of H2O2 per 5 ml of glass distilled water. Carefully remove 15 chamber and rinse slide in water. Air dry for a few minutes, then add 1 drop of Crystal Mount and a cover slip. Bake for 5 min at 85-90° C.

Alternatively, the binding compositions are used to affinity purify or sort out cells expressing the ligand or receptor. See, e.g., Sambrook, et al. or Ausubel et al.

All references cited herein are incorporated herein by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

25 Many modification an variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described herein are offered by way of example only, and the invention is to be limited only by the terms of the appended claims, along with the full scope of the equivalents to which such claims are entitled.

```
SEQUENCE LISTING
      SEQ ID NO: 1 is primate IBICK nucleotide sequence.
      SEQ ID NO: 2 is primate IBICK amino acid sequence.
SEQ ID NO: 3 is primate ILINCK nucleotide sequence.
      SEQ ID NO: 4 is primate ILINCK amino acid sequence.
      SEQ ID NO: 5 is partial rodent CXC-143 nucleotide sequence.
      SEQ ID NO: 6 is partial rodent CXC-143 amino acid sequence.
      SEQ ID NO: 7 is partial alternative rodent CXC-143 nucleotide
10
      sequence.
      SEQ ID NO: 8 is partial alternative rodent CXC-143 amino acid
      sequence.
      SEQ ID NO: 9 is revised partial rodent CXC-143 nucleotide sequence.
      SEQ ID NO: 10 is revised partial rodent CXC-143 amino acid sequence.
15
      SEQ ID NO: 11 is partial rodent MCP243 nucleotide sequence.
      SEQ ID NO: 12 is partial rodent MCP243 amino acid sequence.
      SEQ ID NO: 13 is revised complete rodent MCP243 nucleotide sequence.
      SEQ ID NO: 14 is revised complete rodent MCP243 amino acid sequence.
     SEQ ID NO: 15 is primate R277 nucleotide sequence. SEQ ID NO: 16 is primate R277 amino acid sequence.
20
      SEQ ID NO: 17 is revised primate R277 nucleotide sequence.
      SEQ ID NO: 18 is revised primate R277 amino acid sequence.
      SEQ ID NO: 19 is partial rodent HST01.1 nucleotide sequence.
      SEQ ID NO: 20 is partial rodent HST01.1 amino acid sequence.
25
      SEQ ID NO: 21 is rodent HST01.1 nucleotide sequence.
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      SEQ ID NO: 24 is partial rodent 941D12 amino acid sequence.
      SEQ ID NO: 25 is rodent 941D12 nucleotide sequence.
30
      SEQ ID NO: 26 is rodent 941D12 amino acid sequence.
      (1) GENERAL INFORMATION:
           (i) APPLICANT:
35
            (A)
                  NAME:
                               Schering Corp.
            (B)
                  STREET:
                               2000 Galloping Hill Road
            (C)
                  CITY:
                               Kenilworth
            (D)
                  STATE:
                               New Jersey
            (F)
                  ZIP:
                               07033-0530
40
            (G)
                  TELEPHONE:
                              (908) 298-5056
                  TELEFAX:
            (H)
                               (908) 298-5388
          (ii) TITLE OF INVENTION: Mammalian Chemokines; Receptors;
                  Reagents; Uses
45
         (iii) NUMBER OF SEQUENCES: 26
           (iv) COMPUTER READABLE FORM:
                (A) MEDIUM TYPE: Floppy disk
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                (B) COMPUTER: Macintosh
                (C) OPERATING SYSTEM: Macintosh OS 8.0
                (D) SOFTWARE: Microsoft Word 5.1
          (v) CURRENT APPLICATION DATA:
55
                (A) APPLICATION NUMBER:
```

(B) FILING DATE: 22-JAN-1998

-68-

(C) CLASSIFICATION:

(vi) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: US 60/036,715 5 (B) FILING DATE: 23-JAN-1997 (2) INFORMATION FOR SEQ ID NO:1: (i) SEQUENCE CHARACTERISTICS: 10 (A) LENGTH: 468 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 15 (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: unsure 20 (B) LOCATION: 246 (D) OTHER INFORMATION: /note= "nucleotide 246 designated C, may be A, C, G, or T* (ix) FEATURE: 25 (A) NAME/KEY: CDS (B) LOCATION: 23..307 (ix) FEATURE: (A) NAME/KEY: mat_peptide 30 (B) LOCATION: 86..307 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1: GCAGCAACAG CAAAAAACAA AC ATG AGT GTG AAG GGC ATG GCT ATA GCC TTG 35 Met Ser Val Lys Gly Met Ala Ile Ala Leu -21 -20 GCT GTG ATA TTG TGT GCT ACA GTT GTT CAA GGC TTC CCC ATG TTC AAA 40 100 Ala Val Ile Leu Cys Ala Thr Val Val Gln Gly Phe Pro Met Phe Lys 45 AGA GGA CGC TGT CTT TGC ATA GGC CCT GGG GTA AAA GCA GTG AAA GTG 148 Arg Gly Arg Cys Leu Cys Ile Gly Pro Gly Val Lys Ala Val Lys Val 10 20

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GCA GAT ATT GAG AAA GCC TCC ATA ATG TAC CCA AGT AAC AAC TGT GAC

Ala Asp Ile Glu Lys Ala Ser Ile Met Tyr Pro Ser Asn Asn Cys Asp

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35	(ii) MOLECULE TYPE: protein														
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	Lys Gln Ala Arg Leu Ile Ile Lys Lys Ala Glu Arg Lys Asn Phe														

	60					65				•	70				•		
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5	-	(i)	(<i>E</i> (C	A) LE B) TY C) ST	E CHANGTH: PE: 1 PRANDI	: 50 nucl EDNE	3 ba eic SS:	ase aci sin	pai: .d	rs				•			
10		(ii)	MOL	ECUL	E TYI	PE:	cDN#	A	•								
15 -		(ix)	(A	-	: ME/KE CATIO			.410)							٠.	
20		(ix)	(A		: ME/KE CATIO					e					٠		
	٠	(xi)	SEÇ	UENC	E DES	SCRI	PTIC	ON:	SEQ	ID N	0:3:						
25	GGGG	CTGCA	.GG	AATT	CGGC!	AC	GAG	CTGZ	ĄAGC	TGI	PACTG	CCT	CGC	TGAG	AGG	ATG	AAC
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30	104															C · ATT	ACT
	Val	Ser -20	Glu	Ala	Ala I		Ser -15	Leu	Leu	ı Val	Leu	Ile -10	Leu	Ile	Ile	Thr	
35	152															C, ACC	CC
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•	Ser	Thr	Cys	Cys 15	Leu I	żγs '	Гут	Тут	Glu 20		Val	Leu	Pro	Arg . 25	Arg	Leu	
45	GTG 248	GTG	GGA	TAC	AGA	AAG	GC	ec o	CTC	AAC	TGT	CAC	CTG	CCA	GCZ	А АТС	ATC
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	Phe	Val 45	Thr	Lys	Arg A	Asn A	Arg 50	Glu	Va]	l Cys	Thr	Asn 55	Pro	Asn	Asp	Asp	

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	(ii) MOLECULE TYPE: protein
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20	TAA. 375	\ATA/	AAA		.100	GC	CCT	rtct"	r			тстс	'ACAA	.GC	•		АТАА
25	(2)	INFO	ORMAT	noi	FOR	SEQ	ID 1	NO : 6	:				•		•	•	•
30		,	(i) S	(A)	TYI		10 min	3 am o ac	ino id	: acid:	5	-					
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35		()	ci) S	EQUE	ENCE	DES	RIP	TION	: SE	Q ID	NO:	5 :					
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(2) INFORMATION FOR SEQ ID NO:7: 5 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 374 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single 10 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA 15 (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 3..371 (ix) FEATURE: 20 (A) NAME/KEY: CDS (B) LOCATION: 3..371 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7: 25 GC TCC TGC TCC TCC GCT GTC GCC TCG CGC GTG GAC GGG TCC AAG Ser Cys Cys Ser Ser Ala Val Ala Ser Arg Val Asp Gly Ser Lys 10 30 TGT AAG TGT TCC CGG AAG GGG CCC AAG ATC CGC TAC AGC GAC GTG AAG Cys Lys Cys Ser Arg Lys Gly Pro Lys Ile Arg Tyr Ser Asp Val Lys 35 AAG CTG GAA ATG AAG CCA AAG TAC CCA CAC TGC GAG GAG AAG ATG GTT Lys Leu Glu Met Lys Pro Lys Tyr Pro His Cys Glu Glu Lys Met Val 35 40 40 ATC GTC ACC ACC AAG AGC ATG TCC AGG TAC CGG GGC CAG GAG CAC TGC Ile Val Thr Thr Lys Ser Met Ser Arg Tyr Arg Gly Gln Glu His Cys 45 CTG CAC CCT AAG CTG CAG ACA CCA AAC GCT TCA TCA AGT GGT ACA ATG 239 Leu His Pro Lys Leu Gln Thr Pro Asn Ala Ser Ser Ser Gly Thr Met 50 CCT GGA ACG AGA AGC GCA GGG TCT ACG AAG AAT AGG GTG GAC GAT CAT

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	CAG 374	AT	T A	AAA	АТА	. Až	A.A.	GCC	CT	т т	СТ	TTC	TC.	Α (CAA	GCA	TAA	
10	Gln	Ile	Lys	Ile 115	Lys	Ala	Leu	Ser	Phe 120	Ser	Gln	Ala						
15	(2)	INFO	ORMA?	rion	FOR	SEQ	ID I	мо : 8	:									
		((i) S	(A)		GTH:	: 12	3 am	TICS ino id		5							
20	•			(D)	TOI	POLOG	GY:	line	ar									
		•	•	OLEC			_											
		()	ci) S	EQUE	ENCE	DESC	CRIP	TION	: SE	Q ID	NO:	3:						
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		(i)	(2	QUENC A) LI B) T	ENGT	i: 2	76 b	ase	pair	s								

					RAND:				ngle								
5		(ii)	MOL	ECUL	E TY	PE:	cDN2	A.							-	•	
10		(ix)) NA	: ME/K CATI									٠,			
		(xi)	SEC	WENC	E DE	SCRI	PTIC	ON:	SEQ	ID N	0:9:				·		
15	CTC	CTG	CTG	CTC	CŢC	GC	g Ct	rg	TAC	GCC	TCG	CGC	GTG	GAC	GGG	TCC	AAG
		Leu	Leu	Leu	Leu . 5	Ala	Leu	Ту	r Ala	a Ser 10	Arg	Val	Asp	Gly :	Ser L 15	ys	
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45	(2)	INF	ORMA!	rion	FOR	SEQ	ID :	NO:	10:								
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(ii) MOLECULE TYPE: protein

(A) LENGTH: 91 amino acids(B) TYPE: amino acid

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Leu Leu Leu Leu Leu Leu Ala Leu Tyr Ala Ser Arg Val Asp Gly Ser Lys
5
Cys Lys Cys Ser Arg Lys Gly Pro Lys Ile Arg Tyr Ser Asp Val Lys
30
Lys Leu Glu Met Lys Pro Lys Tyr Pro His Cys Glu Glu Lys Met Val
10
Ile Val Thr Thr Lys Ser Met Ser Arg Tyr Arg Gly Gln Glu His Cys
55
Leu His Pro Lys Leu Gln Ser Thr Lys Arg Phe Ile Lys Trp Tyr Asn
65
Ala Trp Asn Glu Lys Arg Arg Val Tyr Glu Glu
90
20

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 315 base pairs
- 25 (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA

30

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..162

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- AGA AGT TCG GAA CTG AGG GAG AGA ATC AAC AAT ATC CAG TGC CCC ATG 40 48
 Arg Ser Ser Glu Leu Arg Glu Arg Ile Asn Asn Ile Gln Cys Pro Met
- GAA GCT GTG GTT TTC CAG ACC AAG CAG GGT ATG TCT CTC TGT GTA GAC 45 96
 Glu Ala Val Val Phe Gln Thr Lys Gln Gly Met Ser Leu Cys Val Asp 25 30
- CCC ACA CAG AAG TGG GTC AGT GAG TAC ATG GAG ATC CTT GAC CAG AAG

 144

 Pro Thr Gln Lys Trp Val Ser Glu Tyr Met Glu Ile Leu Asp Gln Lys

 35

 40

 45

-	TCT CAA ATT CTG CAG CCT TGAACCTTCA CACCTGAGTT AAGAGACAGC 192 Ser Gln Ile Leu Gln Pro
5	50 CAAAGCTGGA AGTTCTCCCC TAATCTTCTC CAGGCAGAGA GATGTTACAA GCAGATGGTC 252
10	CCTGGGCTGC GTGTTTTCTC ATCCTTGTCT GTTATATGAA CAACTGAAAT AAAAGCTTAC
	ACT 315
15	(2) INFORMATION FOR SEQ ID NO:12:
20	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 54 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: protein
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
	Arg Ser Ser Glu Leu Arg Glu Arg Ile Asn Asn Ile Gln Cys Pro Met 1 5 10 15
30	Glu Ala Val Val Phe Gln Thr Lys Gln Gly Met Ser Leu Cys Val Asp 20 25 30
35	Pro Thr Gln Lys Trp Val Ser Glu Tyr Met Glu Ile Leu Asp Gln Lys 35 40 45
	Ser Gln Ile Leu Gln Pro 50
40	(2) INFORMATION FOR SEQ ID NO:13: (i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 294 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single
45	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA
50	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1291

	<pre>(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 58291</pre>	
5		
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
	ATG AAG ATC TAC GCA GTG CTT CTT TGC CTG CTG CTC ATA GCT GTC 48	CCT
10	Met Lys Ile Tyr Ala Val Leu Leu Cys Leu Leu Leu Ile Ala Val Pro -19 -15 -10 -5	
	GTC AGC CCA GAG AAG CTG ACT GGG CCA GAT AAG GCT CCA GTC ACC	TGC
15	Val Ser Pro Glu Lys Leu Thr Gly Pro Asp Lys Ala Pro Val Thr Cys 1 5 10	
	TGC TTT CAT GTA CTA AAG CTG AAG ATC CCC CTT CGG GTG CTG AAA	AGC
20	Cys Phe His Val Leu Lys Leu Lys Ile Pro Leu Arg Val Leu Lys Ser 15 20 25	
	TAC GAG AGA ATC AAC AAT ATC CAG TGC CCC ATG GAA GCT GTG GTT	TTC
25	Tyr Glu Arg Ile Asn Asn Ile Gln Cys Pro Met Glu Ala Val Val Phe 30 35 40 45	
٠	CAG ACC AAG CAG GGT ATG TCT CTC TGT GTA GAC CCC ACA CAG AAG	TG
30	Gln Thr Lys Gln Gly Met Ser Leu Cys Val Asp Pro Thr Gln Lys Trp 50 55 60	
	GTC AGT GAG TAC ATG GAG ATC CTT GAC CAG AAG TCT CAA ATT CTG	CAG
35	Val Ser Glu Tyr Met Glu Ile Leu Asp Gln Lys Ser Gln Ile Leu Gln 65 70 75	
	CCT	TG
40	294 Pro	

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 97 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

	Met -19	Lys	Ile	Tyr	Ala -15	Val	Leu	Leu	Cys	Leu -10	Leu	Leu	Ile	Ala	-5	Pro	
5	Val	Ser	Pro	Glu 1	Lys	Leu	Thr	Gly 5	Pro	Asp	Lys	Ala	Pro 10	Val	Thr	Cys	
	Cys	Phe 15	His	Val	Leu	Lys	Leu 20	Lys	Ile	Pro	Leu	Arg 25	Val	Leu	Lys	Ser	
10	туr 30		Arg	Ile	Asn	Asn 35	Ile	Gln	Cys	Pro	Met 40	Glu	Ala	Val	Val	Phe 45	
15	Gln	Thr	Lys	Gln	Gly 50	Met	Ser	Leu	Суѕ	Val 55	Asp	Pro	Thr	Gln	Lys 60	Trp	
	Val	Ser	Glu	Туг 65	Met	Glu	Ile	Leu	Asp 70	Gln	Lys	Ser	Gln	Ile 75	Leu	Gln	
20	Pro																
	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:1	5 :								
25		(i	(A) I	CE C	'H: 9	52 b	ase	pair	s						-	
			((C) S	YPE:	DEDN	IESS:	sin	.a .gle							•	
30		(ii			OPOL											÷	
35	I	(i)		(A) 1 (B) 1 (D) (NAME / LOCAT OTHER	TON:	44	7	7: /r	note:	= "nı	ucled	otide	e de:	signa	ated	c,
40		(i:		(A) I	NAME.	TTON	. 48	9	N: /	note	= "n:	ucle	otid	e de	sign	ated	C,
45	1			, C, EATU	G,	or T	Ħ										
50		mav	be A	(B) (D)	T.OCA	TION R IN	: 64 FORM	0		note	= "ח	ucle	otid	e de	sign	ated	A
20			.x) F														
		` -			NAME	1250											

	(B) LOCATION: 480510 (D) OTHER INFORMATION: /note= "possible sequence error; retains reading frame"																
5		(ix	(2	ATURE A) NA B) LC	ME/K			32									
10		(xi)) SE(QUENC	E DE	SCRI	PTIC	n:	SEQ	ID N	0:15	:					
	ATG 48	TAC	ATG	AAT	TT	OAT 7	AG	ic i	AGC	ACA	GCA	TTC	CTC	ACC	TGC	ATT	GCC
15	Met 1	Tyr	Met	Asn	Phe 5	Tyr	Ser	Ser	Thr	Ala 10	Phe	Leu	Thr	Cys	Ile . 15	Ala	
	GTT 96	GAT	CGG	TAT	TTC	G GCT	r GI	T C	GTC	TAC	ССТ	TTG	AAG	TTT	TTI	TTC	ATA
20	Val	Asp	Arg	Tyr 20	Leu	Ala	Val	Val	Туг 25	Pro	Leu	Lys	Phe	Phe 30	Phe	Ile	
	AGG 144	ACA	AGA	AGA	TTT	gc <i>i</i>	A CI	C I	ATG	GTC	AGC	CTG	TCC	ATC	TGG	ATA	TTG
25	Arg	Thr	Arg 35	Arg	Phe	Ala	Leu	Met 40		Ser	Leu	Ser	Ile 45	Trp	Ile :	Leu	
	GAA 192	ACC	ATC	TTC	! AA!	r GC1	r Gī	ec z	ATG	TTG	TGG	GAA	GAT	GAA	ACA	GTT	GTT
30	Glu	Thr 50	Ile	Phe	Asn	Ala	Val 55	Met	Leu	ı Trp	Glu	Asp 60	Glu	Thr	Val '	Val	
	GAA 240	ТАТ	TGC	GAT	GCC	GA/	A AA	G 7	rct	AAT	TTT	ACT	TTA	TGC	TAT	GAC	AAA
35	Glu 65	Tyr	Cys	Asp	Ala	Glu 70	Lys	Ser	Asr	n Phe	Thr 75	Leu	Cys	Tyr	Asp	Lуs 80	
	TAC 288	CCT	TTA	GAG	AA.	A TGO	G CA	A I	ATC	AAC	CTC	AAC	TTG	TTC	AGG	ACG	TGT
40	Tyr	Pro	Leu	Glų	Lys 85	Trp	Gln	Ile	Asr	1 Leu 90	Asn	Leu	Phe	Arg	Thr 95	Cys	
	ACA 336	GGC	TAT	GCA	ATT	י ככי	r T	e c	GTC	ACC	ATC	CTG	ATC	TGC	AAC	c cgg	AAA
45	Thr	Gly	Tyr	Ala 100	Ile	Pro	Leu	Val	Th: 105	: Ile	Leu	Ile	Cys	Asn 110	Arg	Lys	
	GTT 384	TAC	CAA	GCT	GTO	G CGC	C CA	C A	TAA	AAG	GCC	ACG	GAA	AAC	AGO	GAA	AAG
50		Tyr	Gln 115	Ala	Val	Arg	His	Asn 120	-	a Ala	Thr	Glu	Asn 125	Arg	Glu	Lys	
	AGG 432	AGG	ATT	TTA	L AA	A CT	A CI	r r	rtc	AGC	ATC	ACA	GTT	ACT	TTT	GTC	TTA

- Arg Arg Ile Leu Lys Leu Leu Phe Ser Ile Thr Val Thr Phe Val Leu 135 TGT TTT ACC CCC TTC CAG GTG ATG TTG CTG ATC CGC TGC ATT TTA GAG Cys Phe Thr Pro Phe Gln Val Met Leu Leu Ile Arg Cys Ile Leu Glu 155 145 150 CTG CTG TGC ACC TCC GAA GCC CCA CAG CAA TCC GGG AAG CGA ACC TTA 10 528 Leu Leu Cys Thr Ser Glu Ala Pro Gln Gln Ser Gly Lys Arg Thr Leu 165 170 ACA ATG TAT AGA ATC ACG GTT GCC TTA ACC AGT TTA AAA TGT GTT GCT 15 576 Thr Met Tyr Arg Ile Thr Val Ala Leu Thr Ser Leu Lys Cys Val Ala 185 180 GAT CCA ATT CTG TAC TGT TTT GTA ACC GAA ACA GGA AGA TAT GAT ATG 20 624 Asp Pro Ile Leu Tyr Cys Phe Val Thr Glu Thr Gly Arg Tyr Asp Met 200 195 TGG AAT ATA TTA AAA ATC TGC ACT GGG AGG TGT AAT ACA TCA CAA AGA 25 Trp Asn Ile Leu Lys Ile Cys Thr Gly Arg Cys Asn Thr Ser Gln Arg CAA AGA AAA CGC ATA CTT TCT GTG TCT ACA AAA GAT ACT ATG GAA TTA 30 Gln Arg Lys Arg Ile Leu Ser Val Ser Thr Lys Asp Thr Met Glu Leu 235 240 230 225 35 772 Glu Val Leu Glu ATGCATTATT ATATCATCAA GATTACATTT TGAAAAGGAA ATCTAGCATG TGAGGGGACT 40 AAGTGTTCTC AGAGTGATGT TTTAATCCAG TCCAATAAAA ATATCTTAAA ACTGCATTGT 892 ACAGCTCCCT CCCTGCGGTT TTATTAAATG ATGTATATTA AACAAAGATC AATATTTTCA
 - (2) INFORMATION FOR SEQ ID NO:16: 50
 - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 244 amino acids

- (B) TYPE: amino acid
 (D) TOPOLOGY: linear

 DLECULE TYPE: protein
- (ii) MOLECULE TYPE: protein 5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16: Met Tyr Met Asn Phe Tyr Ser Ser Thr Ala Phe Leu Thr Cys Ile Ala 10 Val Asp Arg Tyr Leu Ala Val Val Tyr Pro Leu Lys Phe Phe Phe Ile Arg Thr Arg Arg Phe Ala Leu Met Val Ser Leu Ser Ile Trp Ile Leu 15 Glu Thr Ile Phe Asn Ala Val Met Leu Trp Glu Asp Glu Thr Val Val Glu Tyr Cys Asp Ala Glu Lys Ser Asn Phe Thr Leu Cys Tyr Asp Lys 20 Tyr Pro Leu Glu Lys Trp Gln Ile Asn Leu Asn Leu Phe Arg Thr Cys 90 25 Thr Gly Tyr Ala Ile Pro Leu Val Thr Ile Leu Ile Cys Asn Arg Lys Val Tyr Gln Ala Val Arg His Asn Lys Ala Thr Glu Asn Arg Glu Lys 30 Arg Arg Ile Leu Lys Leu Leu Phe Ser Ile Thr Val Thr Phe Val Leu 135 Cys Phe Thr Pro Phe Gln Val Met Leu Leu Ile Arg Cys Ile Leu Glu 35 155 Leu Leu Cys Thr Ser Glu Ala Pro Gln Gln Ser Gly Lys Arg Thr Leu 170 40 Thr Met Tyr Arg Ile Thr Val Ala Leu Thr Ser Leu Lys Cys Val Ala Asp Pro Ile Leu Tyr Cys Phe Val Thr Glu Thr Gly Arg Tyr Asp Met 45 Trp Asn Ile Leu Lys Ile Cys Thr Gly Arg Cys Asn Thr Ser Gln Arg 215 Gln Arg Lys Arg Ile Leu Ser Val Ser Thr Lys Asp Thr Met Glu Leu 50 235 Glu Val Leu Glu

	(2) INFORMATION FOR SEQ ID NO:17:	
5	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1014 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: cDNA	
15	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 11011	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	_
	ATG AAC AGC ACA TGT ATT GAA GAA CAG CAT GAC CTG GAT CAC TAT TT	G
25	Met Asn Ser Thr Cys Ile Glu Glu Gln His Asp Leu Asp His Tyr Leu 1 15 15	
23	TTT CCC ATT GTT TAC ATC TTT GTG ATT ATA GTC AGC ATT CCA GCC AA	T.
	Phe Pro Ile Val Tyr Ile Phe Val Ile Ile Val Ser Ile Pro Ala Asn 20 25 30	
30	ATT GGA TCT CTG TGT GTG TCT TTC CTG CAA CCC AAG AAG GAA AGT GA	LΑ
	Ile Gly Ser Leu Cys Val Ser Phe Leu Gln Pro Lys Lys Glu Ser Glu 35 40 45	
35	CTA GGA ATT TAC CTC TTC AGT TTG TCA CTA TCA GAT TTA CTC TAT GC	:A
	Leu Gly Ile Tyr Leu Phe Ser Leu Ser Leu Ser Asp Leu Leu Tyr Ala 50 55 60	
40	TTA ACT CTC CCT TTA TGG ATT GAT TAT ACT TGG AAT AAA GAC AAC TG	ΞG
	Leu Thr Leu Pro Leu Trp Ile Asp Tyr Thr Trp Asn Lys Asp Asn Trp 65 70 75 80	
45	ACT TTC TCT CCT GCC TTG TGC AAA GGG AGT GCT TTT CTC ATG TAC A	TC
	288 Thr Phe Ser Pro Ala Leu Cys Lys Gly Ser Ala Phe Leu Met Tyr Met 85 90 95	
50	AAG TTT TAC AGC AGC ACA GCA TTC CTC ACC TGC ATT GCC GTT GAT C	G
	336 Lys Phe Tyr Ser Ser Thr Ala-Phe Leu Thr Cys Ile Ala Val Asp Arg	

	100					105					110						
	384															ACA	AGA
5	Tyr	Leu	Ala 115	Val '	Val '	Tyr 1	Pro	Leu 120	Lys	Phe	Phe	Phe	Leu . 125	Arg '	Thr A	Arg	
,	432															ACC	ATC
10	-	Ile 130	Ala	Leu i	Met '		Ser 135	Leu	Ser	lle	Trp	Ile 140	Leu	Glu '	Thr 1	Ile	
	480															TAT	TGC
15	Phe 145	Asn	Ala	Val		Leu ' 150	Trp	Glu	Asr	Glu	Thr 155	Val	Val	Glu	Tyr (cys 160	
	528															CCT	TTA
20	Asp	Ala	Glu	ГЛЗ	Ser 165	Asn	Phe	Thr	Lev	ı C <u>y</u> s 170	Tyr	Asp	Lys	Tyr	Pro 175	Leu	
	576											-				A GGC	TAT
25	Glu	Lys	Trp	Gln 180	Ile	Asn	Leu	Asr	18:	u Phe 5	Arg	Thr	Cys	Thr 190	G1A	Tyr	
	624									•						TAC	CAA
30	Ala	Ile	Pro 195	Leu	Val	Thr	Ile	200		e Cys	. Asn	Arg	Lys 205	Val	Tyr	Gln	
	672															G AGA	ATC
35	Ala	Val 210		His	Asn	Lys	Ala 215		r Gl	u Asr	n Lys	Glu 220	Lys	Lys	Arg	Ile	
	720															C TTT	ACT
40	11e 225		. Leu	Leu	Val	Ser 230	Ile	e Th	r Va	ıl Th:	r Phe 235	val	Leu	Cys	Pne	240	
·	768	t														T GCT	GTĢ
45	Pro	Phe	e His	; Val	Met 245		Le	ı Il	e Ar	g Cy 25	s Ile O	e Leu	ı Glu	His	A1a 255	vaı	
	816	5													•	A ATG	TAT
50	Asr	n Ph	e Glu	Asp 260		Ser	As	n Se		ly Ly 65	s Ar	g Thi	тух	270	Met	TYY	

							_										
	864															CCA	ATT
	Arg	Ile	Thr 275	Val	Ala	Leu :		Ser 280	Leu	Asn	Cys	Val	Ala 285	Asp	Pro	Ile	
5		TAC	TGT	TTT	GTI	ACC	: GA	A A	.CA	GGA	AGA	TAT	GAT	ATG	TGC	ТАА Э	АТА
	912 Leu	Туг 290	Cys	Phe	Val		Glu 295	Thr	Gly	Arg	Tyr	Asp 300	Met	Trp	Asn	Ile	
10		AAA	TTC	TGC	ACI	GGC	AG	G T	GT	AAT	ACA	TCA	CAA	AGA	CA	A AGA	AAA
	960 Leu 305	Lys	Phe	Суз		Gly . 310	Arg	Cys	Asn		Ser 315	Gln	Arg	Gln	Arg	Lys 320	
15	CGC		CTT	TCI	GTO	TCT	r Ac	A A	AA	GAT	ACT	ATG	GAA	TTA	GA(GTC	CTT
	Arg	Ile	Leu	Ser	Val 325	Ser	Thr	Lys	Asp	330	Met	Glu	Leu	Glu	Val 335	Leu	
20	GAG											•					TAG
	1014	1															
	Glu		•			-											
25											,						
	(2)	INF	ORMA'	noi	FOR	SEQ	ID I	NO:1	8:								
30			(i) :	(B)	LEI TYI	NGTH: PE: a	33° min	7 am	ino id	S: acid	ls						
				(D)	TOI	POLOG	iY:	line	ar								
35				MOLE						EO 11	n NO.	10.					
											NO:						
40	Met 1		Ser	Thr	Суs 5	Ile	Glu	Glu	ı Gl	n His	s Asp O	Leu	. Asp	His	Tyr 15	Leu	
40	Phe	Pro	Ile	Val 20	Тут	Ile	Phe	Va]	L I1 2		e Val	Ser	Ile	Pro 30	Ala	Asn	
45	Ile	Gly	Ser 35		Суз	Val	Ser	Phe 40		u Gl	n Pro	Lys	45	Glu G	Ser	Glu	
	Leu	Gly 50		Tyr	Leu	Phe	Ser 55		ı Se	r Le	u Sei	r Asi	p Lev	ı Lev	Tyr	Ala	
50	Leu 65		c Lev	ı Pro	Leu	Trp 70		a As	р Ту	r Th	r Tr:	p Ası 5	n Lys	s Asp	Asn	Trp 80	
	Thr	. Phe	e Sei	r Pro	Ala	Leu	Cys	· Ly	s Gl	Ly Se	r Ala	a Pho	e Le	ı Met	Tyr	Met	

. 90 85 Lys Phe Tyr Ser Ser Thr Ala Phe Leu Thr Cys Ile Ala Val Asp Arg 105 100 5 Tyr Leu Ala Val Val Tyr Pro Leu Lys Phe Phe Phe Leu Arg Thr Arg 120 Arg Ile Ala Leu Met Val Ser Leu Ser Ile Trp Ile Leu Glu Thr Ile 135 10 Phe Asn Ala Val Met Leu Trp Glu Asp Glu Thr Val Val Glu Tyr Cys 150 155 Asp Ala Glu Lys Ser Asn Phe Thr Leu Cys Tyr Asp Lys Tyr Pro Leu 15 Glu Lys Trp Gln Ile Asn Leu Asn Leu Phe Arg Thr Cys Thr Gly Tyr 185 20 Ala Ile Pro Leu Val Thr Ile Leu Ile Cys Asn Arg Lys Val Tyr Gln 200 Ala Val Arg His Asn Lys Ala Thr Glu Asn Lys Glu Lys Lys Arg Ile 25 Ile Lys Leu Leu Val Ser Ile Thr Val Thr Phe Val Leu Cys Phe Thr Pro Phe His Val Met Leu Leu Ile Arg Cys Ile Leu Glu His Ala Val 30 250 245 Asn Phe Glu Asp His Ser Asn Ser Gly Lys Arg Thr Tyr Thr Met Tyr 265 35 Arg Ile Thr Val Ala Leu Thr Ser Leu Asn Cys Val Ala Asp Pro Ile 280 Leu Tyr Cys Phe Val Thr Glu Thr Gly Arg Tyr Asp Met Trp Asn Ile 40 295 Leu Lys Phe Cys Thr Gly Arg Cys Asn Thr Ser Gln Arg Gln Arg Lys Arg Ile Leu Ser Val Ser Thr Lys Asp Thr Met Glu Leu Glu Val Leu 45 325 Glu 50 (2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

5	(A) LENGTH: 219 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE:														
10	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1219														
15	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19: GAC CGT TAT CTG GCA ATC GTC CAC GCC ACC CAG ATC TAC CGC AGG GA 48	.C													
20	Asp Arg Tyr Leu Ala Ile Val His Ala Thr Gln Ile Tyr Arg Arg Asp 1 5 10 15														
20	CCC CGG GTA CGT GTA GCC CTC ACC TGC ATA GTT GTA TGG GGT CTC TG	T													
	Pro Arg Val Arg Val Ala Leu Thr Cys Ile Val Val Trp Gly Leu Cys 20 25 30														
25	CTG CTC TTT GCC CTC CCA GAT TTC ATC TAC CTA TCA GCC AAC TAC GA	ľΤ													
	144 Leu Leu Phe Ala Leu Pro Asp Phe Ile Tyr Leu Ser Ala Asn Tyr Asp 35 40 45														
30	CAG CGC CTC AAT GCC ACC CAT TGC CAG TAC AAC TTC CCA CAG GTG GC	3T													
	192 Gln Arg Leu Asn Ala Thr His Cys Gln Tyr Asn Phe Pro Gln Val Gly 50 55 60														
35		PA.													
	219 Arg Thr Ala Leu His Val Pro Ser Leu 65 70														
40															
	(2) INFORMATION FOR SEQ ID NO:20:														
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 73 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear														
50	(ii) MOLECULE TYPE: protein														
90	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:														
	Asp Arg Tyr Leu Ala Ile Val His Ala Thr Gln Ile Tyr Arg Arg Asp														

•	1				5			•		10					15			
5	Pro	Arg	Val	Arg 20	Val	Ala	Leu	Thr	Cys 25	Île	Val	Val	Trp	Gly 30	Leu	Cys	5	
כ	Leu	Leu	Phe 35	Ala	Leu	Pro	Asp	Phe 40	Ile	Tyr	Leu	Ser	Ala 45	Asn	Туr	Ası	Þ	
10	Gln	Arg 50		Asn	Ala	Thr	His 55		Gln	Tyr	Asn	Phe 60	Pro	Gln	Val	Gly	7	
	Arg 65	Thr	Ala	Leu	His	Val 70	Pro	Ser	Leu									
15	(2)	INF	ORMA'	rion	FOR	SEQ	ID I	NO:2	1:									
20		(i	() ()	A) L: B) T C) S	CE CI ENGTI YPE: IRANI OPOLO	nuc DEDN	520 1 leic ESS:	base aci sin	pai: d	rs								
		(ii) MO	LECU	LE T	YPE:	cDN.	A		÷								
25		(ix		A) N	E: AME/: OCAT				6									
30		(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:21	:						
35	CTG 60	GAAA	.GGA	AGC.	AGGC	AGC	ACG	AGAC(CTG	ACCO	CCAGO	CAG	CCAC	CAGCO	CGG	AGC	CACC	AGCC
	AAG 107										CGT						GCC	TCG
		М	let T 1	yr L	eu G	lu V	al S	er G	iin A	rg G	ln V	10	eu A	sp A	ia s	er		
40	155										ACC							GGG
45	Asp 15		Ala	. Phe	Leu	. Leu 20		Asn	Ser	Thr	Ser 25		Туг	Asp	Туг		O	
45	203										CCG Pro							TTC
50					. 35	i				40					45	i	٠	CTO
	251								•		Ala					_		

-90-

				50					55	i				60			
	TTC 299	TTG	CTG	GGG	CTG	CT	A GO	GC A	AAT	GGG	GCG	GTG	GCT	GCT	GT	G CTA	CTG
5	Phe	Leu	Leu 65	Gly	Leu	Leu	Gly	Asn 70		Ala	Val	Ala	Ala 75	Val	Leu	Leu	
	AGT 347		CGC	ACT	GCC	CT	G AC	SC 2	AGC	ACG	GAC	ACC.	TTC	CTG	CTO	C CAC	CTG
10	Ser	Gln 80	Arg	Thr	Ala	Leu	Ser 85	Ser	Thr	Asp	Thr	Phe 90	Leu	Leu	His	Leu	
	395															G GCA	GTG
15	Ala 95	Val	Ala	Asp		Leu 100	Leu	Val	Leu	Thr	Leu 105	Pro	Leu	Trp	Ala	Val 110	
	443													-		A GTG	GCA
20	Asp	Ala	Ala	Val	Gln 115	Trp	Val	Phe	Gly	Pro 120	Gly	Leu	Cys		Val 125	Ala	
	491															G CTG	GCT
25	Gly	Ala	Leu	Phe 130	Asn	Ile	Asn	Phe	135		Gly	Ala	Phe	Leu 140	Leu	Ala	
	539															C CAG	ATC
30	Cys	Ile	Ser 145	Phe	Asp	Arg	Tyr	150		lle	Val	His	Ala 155	Thr	GIn	IIe	
	587															A GTT	GTA
35	Tyr	Arg 160	Arg	Asp	Pro	Arg	Val 165	Arg	r Val	. Ala	Leu	170	Cys	Ile	Val	Val	
	635										CCA					C CTA	TCA
40	Trp 175	Gly	Leu	Cys	Leu	Leu 180	Phe	Ala	ı Leı	ı Pro	185	Phe	Ile	Tyr	Leu	Ser 190	٠:
	683															C AAC	TTC
45	Ala	Asn	Tyr	Asp	Gln 195	Arg	Leu	Asr	n Ala	200		Cys	Gln	Tyr	Asn 205	Phe	
	731			,												G GCT	GGT
50	Pro	Gln	Val	Gly 210	Arg	Thr	Ala	Lev	21!		. Leu	Gln	Leu	Val 220	Ala	Gly	

	TTC 779	CTG	CTG	CCC	CTI	ст	G G	TC	ATG	GCC	TAC	TGC	TAT	GCC	CA	r ATC	CTA
		Leu	Leu 225	Pro	Leu	Leu	Val	Met 230		a Tyr	Cys	Tyr	Ala 235	His	Ile	Leu	
5	GCT 827	GTT	CTG	CTG	GTC	тс	C A	GA	GGC	CAG	AGG	CGT	TTT	CGA	. GC	r atg	AGG
		Val 240	Leu	Leu	Val	Ser	Arg 245		y Gli	n Arg	Arg	Phe 250	Arg	Ala	Met	Arg	
10	CTA 875	GTG	GTA	GTG	GTC	GT	G G	CA	GCC	TTT	GCT	GTC	TGC	TGG	AC	e ece	TAT
		Val	Val	Val	Val	Val 260	Ala	Ala	a Phe	e Ala	Val 265	Cys	Trp	Thr	Pro	Tyr 270	
15		CTG	GTG	GTG	CT?	A GT	'G G	АТ	ATC	CTC	ATG	GAT	GTG	GGA	, GT	T TTG	GCC
	923 His	Leu	Val	Val	Leu 275	Val	Asp	Il	e Le	u Met 280		Val	Gly	Val	Leu 285	Ala	
20		AAC	TGT	· GGT	. CG	A AA	A A	.GC	CAC	GTG	GAT	GTG	GCC	AAC	TC.	A GTO	ACC
	971 Arg	Asn	Cys	Gly 290	Arg	Lys	Ser	Hi	s Va 29	l Asp 5	Val	Ala	Lys	Ser 300	Val	Thr	
25			ATG	GGG	TA	C AT	rg c	AC	TGC	TGC	CTC	AAT	CCG	CTC	CT	C TAT	r GCC
	101: Ser	Gly	Met 305	Gly	Tyr	Met	His	Cy		s Leu	Asn	Pro	Leu 315	Leu	Tyr	Ala	
30			GG#	GTO	G AA	G T7	rc A	AGA	GAG	AAA	ATG	TGG	ATG	TT	3 TT	C ACC	G CGC
	106 Phe	7 Val 320	Gly	Val	Lys	Phe	Arc 325		u Ly	s Met	Trp	Met 330	Leu	Phe	Thr	Arg	
35		GGC	: CGC	TC'	r ga	C CI	AG #	AGA	GGG	CCC	CAG	CGG	CAG	cc	G TC	A TC	r TCA
	111 Leu 335	Gly	Arg	Ser	Asp	Gln 340	Arg	g Gļ	y Pr	o Glr	Arg 345		Pro	Ser	Ser	Ser 350	
40	CGG	AGA	GAZ	A TC	A TC	C TO	GG 7	rct	GAG	ACA	ACT	GAG	GCC	TC	C TA	CTO	G GGC
	116 Arg		Glu	Ser	Ser 355	Trp	Se	r Gl	u Th	r Thi		Ala	Ser	Tyr	Leu 365	Gly	
45	TTG 121 Leu	.6	TTAAT	CTGG	Α	CTG	GAAC	TGT	A	GCCT(GCGCA	. с	SCCCA	AGTC	С	TAACA	\CACT(
50			TTG	TCC	TCCT	rgt	AGT	rtgg	GCTA	. GCI	'CGAA	CTT	ACC	CGTA	ACT	TTGC	rgcca
	127	6															

	GATGCACTGA	CAGCTCAGCA	TATATCCAG	G TCTCCTGAG.	A ATCAATTTC	A GCAACAAGGA
5	CAACACCATT	ACTGTGCCTT	AGCTGCCAT	G CCCTATCTT	G CTGTTTAG	A ACTAGCTGCC
	TGGAGCCCCA	CCGCCCTACT	AAATTAGCA	A GTAGAACTC	A GCCATCCCT	G TGTGAGAAGA
10	GGGAGAGGCA 1516	AATAGCACAG	AGGGCCAGG	C GTTGTCAGC	A CTGAATGTG	C CCATCTCAGT
	ATCTCAATAT 1576	TTGCCCAATT	TTATTTCT	AG AAACCTCAC	TAAACTTTC	A ATAAACAAGG
15	TAATGAGGGA 1620	AAAAA	AAAAA	AAAAAAAA A.	AAAAAA	AAA AAAA
20	(2) INFORM	ATION FOR S	EQ ID NO:22	:		•
20		SEQUENCE C	HARACTERIST	ICS:		
	• • •	(A) LENG	TH: 367 ami : amino aci	no acids	,	
25		(D) TOPO	LOGY: linea	ir		
	(ii)	MOLECULE T	YPE: protei	.n		•
30				SEQ ID NO:2		
30	Met Tyr Le 1	u Glu Val S 5	er Glu Arg	Gln Val Leu 10	Asp Ala Ser	Asp Phe 15
35	Ala Phe Le	eu Leu Glu A 20	sn Ser Thr	Ser Pro Tyr 25	Asp Tyr Gly 30	Glu Asn
•		sp Phe Ser A	Asp Ser Pro 40	Pro Cys Pro	Gln Asp Phe 45	Ser Leu
40	Asn Phe As	sp Arg Thr I	he Leu Pro 55	Ala Leu Tyr	Ser Leu Leu 60	Phe Leu
45	Leu Gly L	eu Leu Gly i	Asn Gly Ala 70	Val Ala Ala 75	Val Leu Leu	Ser Gln 80
45	Arg Thr A	la Leu Ser 85	Ser Thr Asp	Thr Phe Leu 90	Leu His Leu	Ala Val 95
50		al Leu Leu 100	Val Leu Thr	Leu Pro Leu 105	Trp Ala Val	Asp Ala
	31 - W-1 C	in Trp Val	Phe Gly Pro	Gly Leu Cys	Lys Val Ala	Gly Ala

]	Ĺeu	Phe 130	Asn	Ile	Asn	Phe	Tyr 135	Ala	Gly	Ala	Phe	Leu 140	Leu	Ala	Суѕ	Ile
5		Ser 145	Phe	Asp	Arg	Tyr	Leu 150	Ser	Ile	Val	His	Ala 155	Thr	Gln	Ile	Tyr	Arg 160
10		Arg	Asp	Pro	Arg	Val 165	Arg	Val	Ala	Leu	Thr 170	Cys	Ile	Val	Val	Trp 175	Gly
	• ;	Leu	Cys	Leu	Leu 180	Phe	Ala	Leu	Pro	Asp 185	Phe	Ile	Tyr	Leu	Ser 190	Ala	Asn
15		Tyr	Asp	Gln 195	Arg	Leu	Asn	Ala	Thr 200	His	Cys	Gln	Tyr	Asn 205	Phe.	Pro	Gln
	,	Val	Gly 210	Arg	Thr	Ala	Leu	Arg 215	Val	Leu	Gln	Leu	Val 220	Ala	Gly	Phe	Leu
20		225				Val	230	ē			•	235					240
25		Leu	Leu	Val	Ser	Arg 245	Gly	Gln	Arg	Arg	Phe 250	Arg	Ala	Met	Arg	Leu 255	Val
					260	Ala				265			-		270		
3() .			275					280					285			Asn
			290					295					300				Gly
3 5		305					310					315					Val 320
4	0					325					330			•		335	Gly
				•	340					345					350		Arg
4	5	Glu	Ser	Ser 355		Ser	Glu	Thr	Thr 360		Ala	Ser	Tyr	Leu 365	Gly	Leu	•
		(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:2	3:	•						

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 581 base pairs

(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 3..581 (ix) FEATURE: 10 (A) NAME/KEY: unsure (B) LOCATION: 169..521 (D) OTHER INFORMATION: /note= "nucleotides 169, 178, 217, 287, 290, 382, 386, 395, 411, 484, 512, 515, 517, 521 each designated C; may be A, C, G, or T* 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23: AG ATG CAG ACT TTA GTG AGC ACA CTT CAC TCT GGA ACA AAG CTA CTG 20 Met Gln Thr Leu Val Ser Thr Leu His Ser Gly Thr Lys Leu Leu . 5 10 GGC TTC TCT GAT GCC ATG GAT GAT GGG CAT CAA GAG TCA ACT CTG 25 Gly Phe Ser Ser Asp Ala Met Asp Asp Gly His Gln Glu Ser Thr Leu 20 TAC GAT GGG CAC TAC GAG GGA GAT TTC TGG CTC TTC AAC AAT TCC AGT 30 Tyr Asp Gly His Tyr Glu Gly Asp Phe Trp Leu Phe Asn Asn Ser Ser GAT AAC AGC CAG GAG AAC AAA CGC TCC CTA AAG TCC AAG GAG GTC TTT 35 Asp Asn Ser Gln Glu Asn Lys Arg Ser Leu Lys Ser Lys Glu Val Phe 50 55 TTG CCC TGT GTG TAC CTG GTA GTG TCT GTC TTT GGA CTG CTA GGA AAC 40 Leu Pro Cys Val Tyr Leu Val Val Ser Val Phe Gly Leu Leu Gly Asn TCC CTG GTT CTG ATT ATA TAC ATT TTC TAC CAA AAG CTG AGG ACT CTC 45 Ser Leu Val Leu Ile Ile Tyr Ile Phe Tyr Gln Lys Leu Arg Thr Leu ACC GAT GTG TTT CTG CTG AAC TTG CCC CTG GCT GAC CTG GTG TTT GTC 50 335 Thr Asp Val Phe Leu Leu Asn Leu Pro Leu Ala Asp Leu Val Phe Val

105

	TGT 383	ACT	CTG	ccc	TTT	TGG	GC	C T	AT (GCA	AGC	ACC	TAT	GAG	TG	G GTC	TCT
		Thr	Leu	Pro 115	Phe ?	Trp /	Ala	Tyr	Ala 120	Ser	Thr	Tyr	Glu	Trp 125	Val	Ser	
5		ACA	GTC	ATC	TTC	AAA	AC	т с	rt (CGA	CGC	ATG	TTA	TAC	AA'	r gaa	TTC
	431 Gly	Thr	Val 130	Ile	Phe l	Lys :		Leu 135	Arg	Arg	Met	Leu	Tyr 140	Asn	Glu	Phe	
10																	
	479															G TTT	CAT
1 =	Tyr	Val 145	Phe	Met	Leu '		Leu 150	Thr	Суѕ	lle	Thr	155		ren	Pne	nis	
15	TGT 527	ACT	GGT	CCA	GCT	ACC	C AA	.G G	CC	TTC	AAC	CGC	CAC	GCT	' AA	C TGG	AAA
		Thr	Gly	Pro		Thr 1 165	Lys	Ņlа	P'ne	Asn	Arg 170	His	Ala	Asn	Trp	Lys 175	
20		CTT	GGG	GCC	TAA :	TC?	A TT	T G	СТ	TGC	TCA	TTT	GGT	TGT	CT	C CCT	GTI
	575 Asn	Leu	Gly	Ala		Ser	Phe	Ala	Cys	Ser 185	Phe	Gly	Суѕ	Leu	Pro 190	Val	
25				•	180					100					170		
	GGG 581						٠				•						TTC
	Gly	Phe											ę.	•			
30																	
	(2)	INF	ORMA'	rion	FOR	SEQ	ID N	10:2	4:								
35			(i) :	(A)	ENCE LEN TYP	GTH: E: a	193 mino	am.	ino id		ls						
40		(ii)	MOLE	CULE	TYPE	: ·pi	rote	in							•	
		(xi)	SEQUI	ENCE	DESC	RIP	rion	: SE	EQ II	NO:	24:					
4 ===	Met 1		Thr	Leu	Val 5	Ser	Thr	Leu	His	S Sei	c Gly O	Thr	ГÀЗ	Leu	Leu 15	Gly	
45	Phe	Ser	Ser	Asp 20	Ala	Met	Asp	Asp	G1 ₃		s Gln	Glu	Ser	Thr 30		Tyr	-
50	Asp	Gly	His		Glu	Gly	Asp	Phe 40		p Le	u Phe	a Asr	Asn 45		Ser	Asp	
	Asn	Ser	Gln	Glu	Asn	Lys	Arg	Ser	Le	u Ly:	s Ser	Lys	Glu	Val	Phe	Leu	

	Pro 65	Cys	Val	Tyr	Leu	Val 70	Val	Ser	Val	Phe	Gly 75	Leu	Leu	Gly	Asn	Ser 80
5	Leu	Val	Leu	Ile	Ile 85	Тут	Ile	Phe	Tyr	Gln 90	Lys	Leu	Arg	Thr	Leu 95	Thr
10	Asp	Val	Phe	Leu 100	Leu	Asn	Leu	Pro	Leu 105	Ala	Asp	Leu	Val	Phe 110	Val	Cys
10	Thr	Leu	Pro 115	Phe	Trp	Ala	Tyr	Ala 120	Ser	Thr	Tyr	Glu	Trp 125	Val	Ser	Gly
15	Thr	Val 130	Ile	Phe	Lys	Thr	Leu 135	Arg	Arg	Met	Leu	Туг 140	Asn	Glu	Phe	Tyr
-	Val 145	Phe	Met	Leu	Thr	Leu 150	Thr	Суз	Ile	Thr	Val 155	Asp	Leu	Phe	His	Cys 160
20	Thr	Gly	Pro	Ala	Thr 165	Lys	Ala	Phe	Asn	Arg 170	His	Ala	Asn	Trp	Lys 175	Asn
25	Leu	Gly	Ala	Asn 180	Ser	Phe	Ala	Cys	Ser 185	Phe	Gly	Суз	Leu	Pro 190	Val	Gly
23	Phe															
30	(2)) SE(QUENC	FOR CE CI ENGTI YPE:	HARA	CTER	ISTI base	CS: pai:	rs		٠				
35			-		TRANI OPOL				gle					•		
		(ii) MO	LECU	LE T	YPE:	cDN.	A				•				
40		(ix	•	A) N.	E: AME/ OCAT											
45	c	·	(A) N B) L D) O	AME/ OCAT	ION:	942				"nu	cleo	tide	942	des	ignated
50		(ix	Ċ	A) N B) L	AME/ OCAT	ION:	141	2			"nu	cleo	tide	s 14	12 a	nd 1422

each designated C, may be A, C, G, or T*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25: GATGCC ATG GAT GAT GGG CAT CAA GAG TCA GCT CTG TAC GAT GGG_ CAC 10 Met Asp Asp Gly His Gln Glu Ser Ala Leu Tyr Asp Gly His TAC GAG GGA GAT TTC TGG CTC TTC AAC AAT TCC AGT GAT AAC AGC CAG 15 Tyr Glu Gly Asp Phe Trp Leu Phe Asn Asn Ser Ser Asp Asn Ser Gln GAG AAC AAA CGC TTC CTA AAG TTC AAG GAG GTC TTT TTG CCC TGT GTG 20 204 Glu Asn Lys Arg Phe Leu Lys Phe Lys Glu Val Phe Leu Pro Cys Val TAC CTG GTA GTG TTT GTC TTT GGA CTG CTA GGA AAC TCC CTG GTT CTG 25 252 Tyr Leu Val Val Phe Val Phe Gly Leu Leu Gly Asn Ser Leu Val Leu ATT ATA TAC ATT TTC TAC CAG AAG CTG AGG ACT CTG ACA GAT GTG TTT 30 Ile Ile Tyr Ile Phe Tyr Gln Lys Leu Arg Thr Leu Thr Asp Val Phe CTG CTG AAC TTG CCC CTG GCT GAC CTG GTG TTT GTC TGT ACT CTG CCC 35. Leu Leu Asn Leu Pro Leu Ala Asp Leu Val Phe Val Cys Thr Leu Pro ឧ೧ TTT TGG GCC TAT GCA GGC ACC TAT GAG TGG GTC TTT GGC ACA GTC ATG 40 Phe Trp Ala Tyr Ala Gly Thr Tyr Glu Trp Val Phe Gly Thr Val Met 105 TGC AAA ACT CTT CGA GGC ATG TAT ACA ATG AAC TTC TAC GTG TCC ATG 45 Cys Lys Thr Leu Arg Gly Met Tyr Thr Met Asn Phe Tyr Val Ser Met CTC ACT CTC ACC TGC ATC ACA GTG GAT CGT TTC ATT GTA GTG GTC CAG 50 492 Leu Thr Leu Thr Cys Ile Thr Val Asp Arg Phe Ile Val Val Gln

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	GCT 540	ACC	AAG	GCC	C TT	C A	AC C	GG	CAG	GCT	AAG	TGG	AAG	ATO	TG	G GGC	CAA
5	Ala	Thr	Lys 145	Ala	Phe	Asn	Arg	Gl 15		a Lys	Trp	Lys	Ile 155	Trp	Gly	Gln	
5	GTC 588	ATT	TGC	TTC	G CT	C AT	т т	GG	GTG	GTC	TCC	CTG	TTG	GT?	r TC	T TTG	CCA
10	Val	Ile 160	Cys	Leu	Leu	Ile	Trp 165		l Va	l Ser	Leu	Leu 170	Val	Ser	Leu	Pro	
10	CAG 636	ATC	ATC	TAT	r GG	C CA	AT G	TT	CAA	GAT	ATT	GAC	AAG	CTI	TA T	'C TGT	CAG
15	Gln 175	Ile	Ile	Tyr	Gly	His 180	Val	G1	n Asj	p Ile	Asp 185	Lys	Leu	Ile	Суѕ	Gln 190	
13	TAC 684	CAC	AGT	GAC	G GA	G AI	T AT	CC	ACT	ATG	GTT	CTT	GTT	ÄTA	A CA	G ATG	ACT
20	Туr	His	Ser	Glu	Gl·u 195	Ile	Ser	Th	r Me	t Val 200		Val	Ile	Gln	Met 205	Thr	
20	CTG 732	GGG	TTC	TTC	CT	g co	CA T	TG	стс	ACT	ATG	ĄTT	CTG	TGC	т ТА	C TĊA	GGC
	Leu	Gly	Phe	Phe 210	Leu	Pro	Leu	Le	u Th:	r Met 5	Ile	Leu	Cys	Tyr 220	Ser	Gly	
25	ATT 780	ATC	AAG	ACC	C TT	G CI	T C	АТ	GCT	CGA	AAC	TTC	CAG	AAC	G CA	.С ААА	TCT
2.0	Ile	Ile	Lys 225	Thr	Leu	Leu	His	23		g Asn	Phe	Gln	Lys 235	His	Lys	Ser	
30	CTA 828	AAG	ATC	ATO	C TT	c ci	T G	TA	GTG	GCT	GTG _.	TTC	CTG	CTC	AC	C CAG	ACA
35	Leu	Lys 240	Ile	Ile	Phe	Leu	Val 245		l Al	a Val	Phe	Leu 250	Leu	Thr	Gln	Thr	
33	CCC 876	ŢTC	AAC	CTI	r GC	C AT	G I	'TA	ATC	CAA	AGT	ACA	AGC	TGC	G GA	G TAC	TAT
<u>4</u> 0	Pro 255	Phe	Asn	Leu	Ala	Met 260	Leu	Il	e Gl	n Ser	Thr 265	Ser	Trp	Glu	Tyr	Tyr 270	
± U	ACC 924	ATA	ACC	AGO	TT	T A.	AG T	'ΑΤ	GCC	ATC	GTA	GTG	ACA	GAG	G GC	т ата	GCA
45	Thr	Ile	Thr	Ser	Phe 275	Lys	Tyr	Al	a Il	e Val 280		Thr	Glu	Ala	Ile 285	Ala	
# 3	TAC 972	TTT	CCG	GGG	TT	G CI	гс т	TA	ACC	CTG	TAC	TTT	ATG	CCI	r TT	G TTG	GCT
= 0	Tyr	Phe	Pro	Gly 290	Leu	Leu	Leu	Th	r Le	u Tyr 5	Phe	Met	Pro	Leu 300	Leu	Ala	
50	TAA	AGTTO	CCG	GAAG	BAACO	GTC	TGG.	AAA(CTTA	TGA	AGGAT	'AT	cggc	TGCC	TC	TCTCAC	CTGG

•	GAGTO	TCA	AG	TCAA	TGGA	AG	TCTT	CTGA	GG Z	AC AG '	TTCC.	AA (GACT:	rgtt(CT (GCCTC	CCACA
5	ATGTA 1152	GAG.	AC	CACC	AGTA	TG	TTCC.	AATT	GT i	AGTA	GGCC'	rt (GCCA	CACT"	ra (GAGAA	GTTAA
	TAACA 1212	GAA'	тт	CTAG	GAGC	AT	GGCT	GTAT	CA '	rttg(GATG	CA Z	ACAA	SAAA	AG (CTTTG	CTTAT
10	AGCAT	GTG	GA	GTAT	CATG	GA ·	GAAA	GTCA	CT (GAAC	ACCA'	rg (GCTG	STAC	AC A	AAAAC	TTCTC
15	AGATA	TAA.	AT	ATAC	CCTA	TT	СТТА	АТАТ	CT I	AAGC	CTAA'	rg (CTCA	AAGG	AG 2	AATGA	GTTAT
13	CCTTG 1392	GAGA	TT	TTGA	AGCA	CT	TTCT	СТСТ	TT (CATC	CCTC	CA Z	AGAA	ATGC!	rg 2	AAATC	AAGGT
20	CCATC 1452	GACG	GT	TAAC	TCCT	AC	АААТ	TCTT	cc i	ATTT(CCTC	et '	TTTT.	PACC	CA Z	AATTT	TTGGG
	CCCTA 1475	AAA	AT				٠		TTI	GAAA	AAA						CCT
25	(2) I	NFO	RMA!	rion	FOR	SEQ	ID N	10:26	5:						٠		
30		(i) :	(B)	LEN TYP	IGTH:		ami aci	.no a .d								
		(i	i) !	MOLEC	CULE	TYPE	E: pr	otei	in								
35		(x	i) :	SEQUE	ENCE	DESC	RIPT	NOI?	SEC	O ID	NO:2	6:					
	Met A	qeA	Asp	Gly	His 5	Gln	Glu	Ser	Ala	Leu 10	Tyr	Asp	Gly	His	Tyr 15	Glu	
40	Gly A	Asp	Phe	Trp	Leu	Phe	Asn	Asn	Ser 25	Ser	Asp	Asn	Ser	Gln 30	Glu	Asn	
45	Lys A	Arg	Phe 35	Leu	Lys	Phe	Lys	Glu 40	Val	Phe	Leu	Pro	Cys 45	Val	Tyr	Leu	
	Val V	Val 50	Phe	Val	Phe	Gly	Leu 55	Leu	Gly	Asn	Ser	Leu 60	Val	Leu	Ile	Ile	
50	Tyr :	Ile	Phe	Tyr	Gln	Lys 70	Leu	Arg	Thr	Leu	Thr 75	Asp	Val	Phe	Leu	Leu 80	
	Asn 1	Leu	Pro	Leu	Ala 85	Asp	Leu	Val	Phe	Val 90	Cys	Thr	Leu	Pro	Phe 95	Trp	

				100					103		Gly					
5			115					120			Tyr					
	Leu	Thr	Суѕ	Ile	Thr	Val	Asp 135	Arg	Phe	Ile	Val	Val 140	Val	Gln	Ala	Thr
10	145					150					11e					
15	Суѕ	Leu	Leu	lle	Trp	Val	Val	. Sei	Lev	170	ı Val	Ser	Leu	Pro	Gln 175	Ile
	Ile	туг	Gly	/ His	; Val	Glr	n Asp) Ile	e Ası	Lys 5	s Lev	ılle	: Cys	190	Tyr)	His
20			19	5				20	U							Gly
		21	0				21	כ								e Ile
25	22	5				23	U					•				1 Lys 240
30	Il	e Il	e Ph	e Le	u Va 24	11 Va 15	1 Al	a Va	al Pi	ie Le 25	eu L∈ 50	u Th	r Gl	n Th	r Pr 25	o Phe 5
	As	n Le	eu Al	La Me 26	t Le	eu Il	Le G	in S	er Ti	nr S 65	er Ti	rp Gl	.и Ту	r Ty 27	r Th	r Ile
35	Tì	ır Se	er Pl	he Ly 75	ys T	yr A	la I	le V 2	al V 80	al T	hr G	lu A	la II 28	le A: 85	la Ty	r Phe
	P:		łу L 90	eu L	eu L	eu T	hr L 2	eu T 95	yr P	he M	iet P	ro L 3	eu L	eu A	la	
40																

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WHAT IS CLAIMED IS:

- 1. A substantially pure or isolated rodent CXC-143 protein or peptide which exhibits at least about 85% sequence identity over a length of at least about 12 amino acids to SEQ ID NO: 6, 8, or 10.
 - 2. A substantially pure or isolated rodent MCP243 protein or peptide which exhibits at least about 85% sequence identity over a length of at least about 12 amino acids to SEQ ID NO: SEQ ID NO: 12 or 14.
 - 3. A substantially pure or isolated primate R277 protein or peptide which exhibits at least about 85% sequence identity over a length of at least about 12 amino acids to SEQ ID NO: 16 or 18.
- 4. A substantially pure or isolated rodent HST01.1 protein or peptide which exhibits at least about 85% sequence identity over a length of at least about 12 amino acids to SEQ ID NO: 20 or 22.
- 5. A substantially pure or isolated rodent 941D12 protein or peptide which exhibits at least about 85% sequence identity over a length of at least about 12 amino acids to SEQ ID NO: 24 or 26.
- 6. A fusion protein comprising the protein or peptide of any of claims 1-5.
 - 7. A binding compound which specifically binds to the protein or peptide of any of claims 1-5.
- 30 8. The binding compound of claim 7 which is an antibody or antibody fragment.
 - A nucleic acid encoding the protein or peptide of any of claims
 1-5.
 - 10. An expression vector comprising the nucleic acid of claim 9.

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- A host cell comprising the vector of claim 10.
- 12. A process for recombinantly producing a polypeptide comprising culturing the host cell of claim 11 under conditions in which the polypeptide is expressed.
 - 13. A method of producing a ligand:receptor complex, comprising contacting:
- a) a substantially pure primate IBICK protein with a G protein coupled receptor;
 - b) a rodent CXC-143 protein or peptide of claim 1 with a G protein coupled receptor;
 - c) a rodent MCP243 protein or peptide of claim 2 with a G protein coupled receptor;
 - d) a primate R277 protein or peptide of claim 3 with a chemokine or ligand; or
 - e) a rodent HST01.1 protein or peptide of claim 4 with a chemokine or ligand; or
- 20 f) a rodent 941D12 protein or peptide of claim 5 with a chemokine or ligand; thereby allowing said complex to form.
 - 14. The method of Claim 13, wherein:
- a) said complex results in a Ca++ flux or cell chemotaxis;
 - b) said G protein coupled receptor is on a cell;
 - c) said complex results in a physiological change in a cell expressing said receptor or protein;
 - d) said primate R277 or murine HST01.1 or 941D12 protein is on a cell;
 - e) said contacting is with a sample comprising a chemical antagonist to block production of said complex; or
 - f) said contacting allows quantitative detection of said ligand.
- 35 15. A method of:

- a) blocking an inflammatory response mediated by IL-10, comprising treating a cell with an antagonist of an ILINCK chemokine; or
- b) inducing an inflammatory response mediated by IL-10, comprising contacting a cell with an ILINCK chemokine.